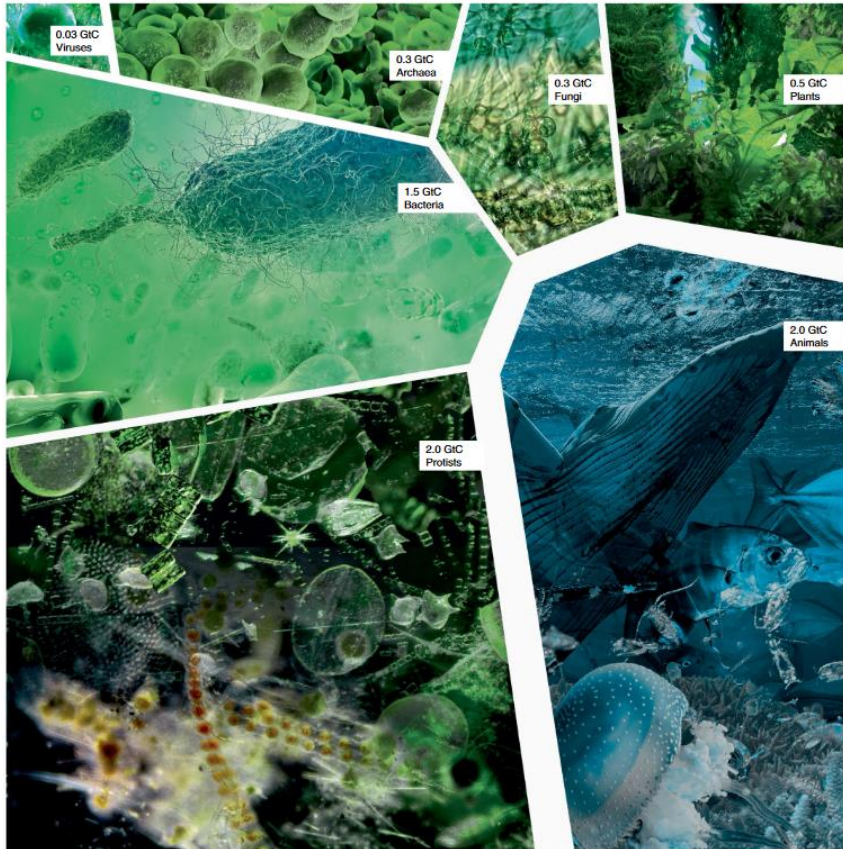


# Shifts in phytoplankton and bacterioplankton communities induced by simulated freshwater intrusion near ice shelves in the Pacific sector of the Southern Ocean

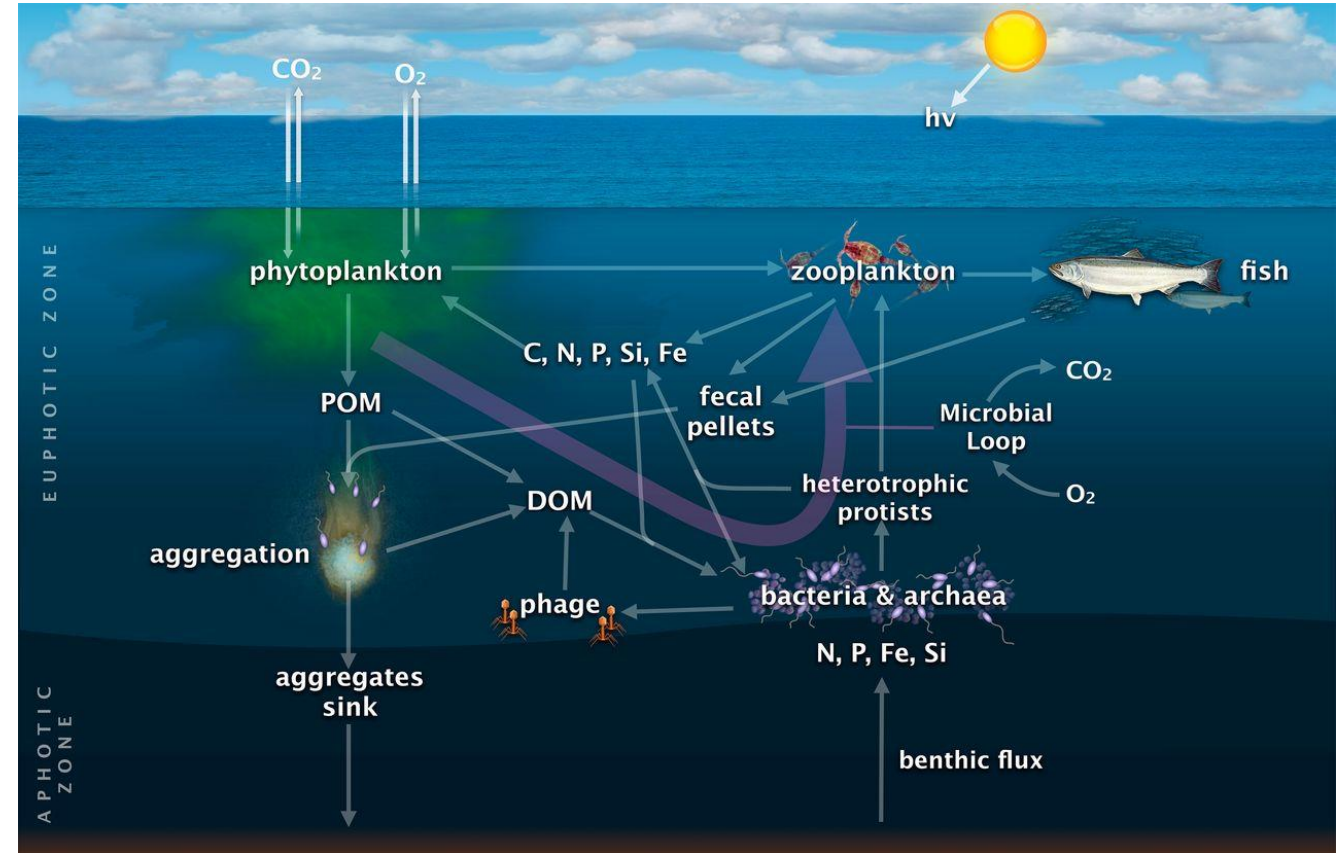
**Jaeho Choi**

Ph. D. student  
Seoul National University  
Korea Polar Research Institute

# Marine microbial community in ocean environment



Abreu *et al.* (2022)

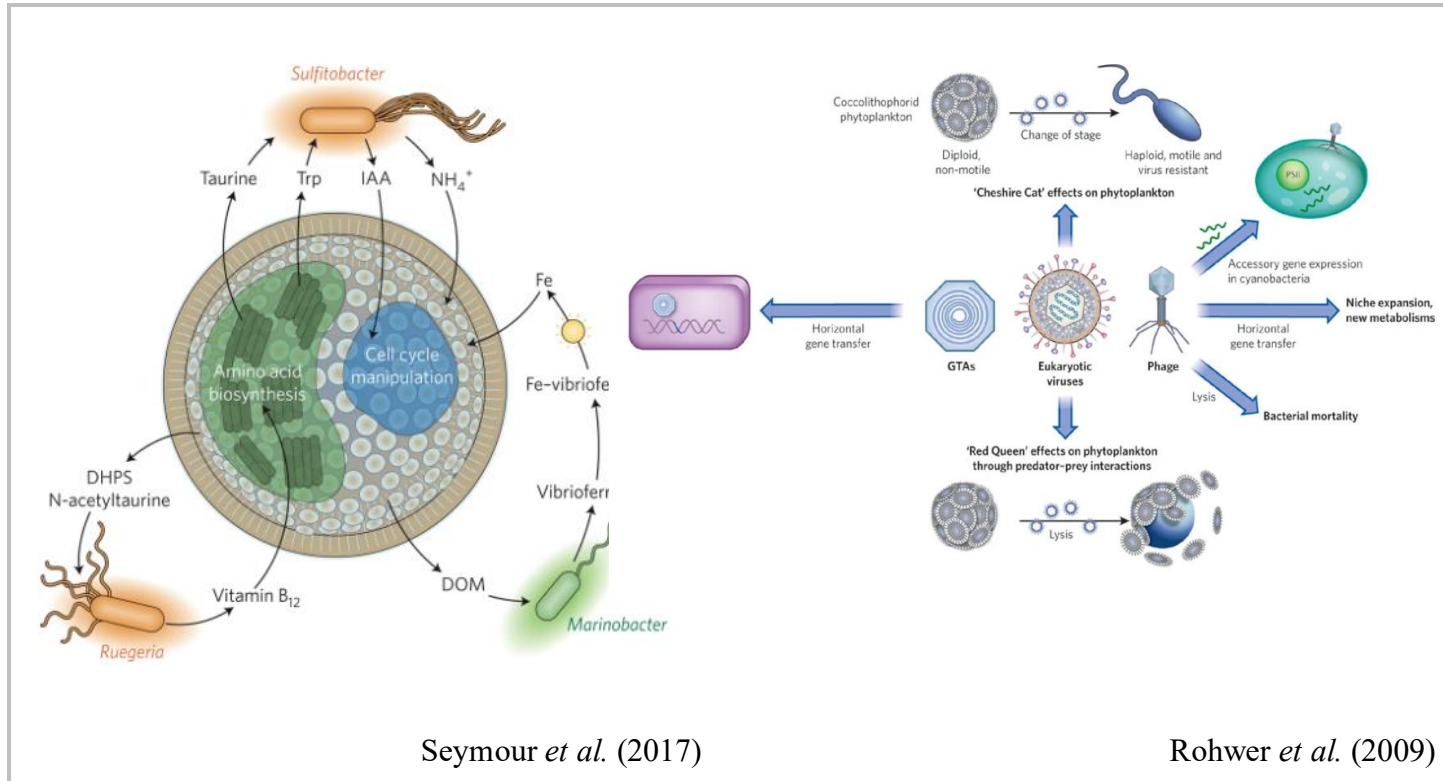


Worden *et al.* (2015)

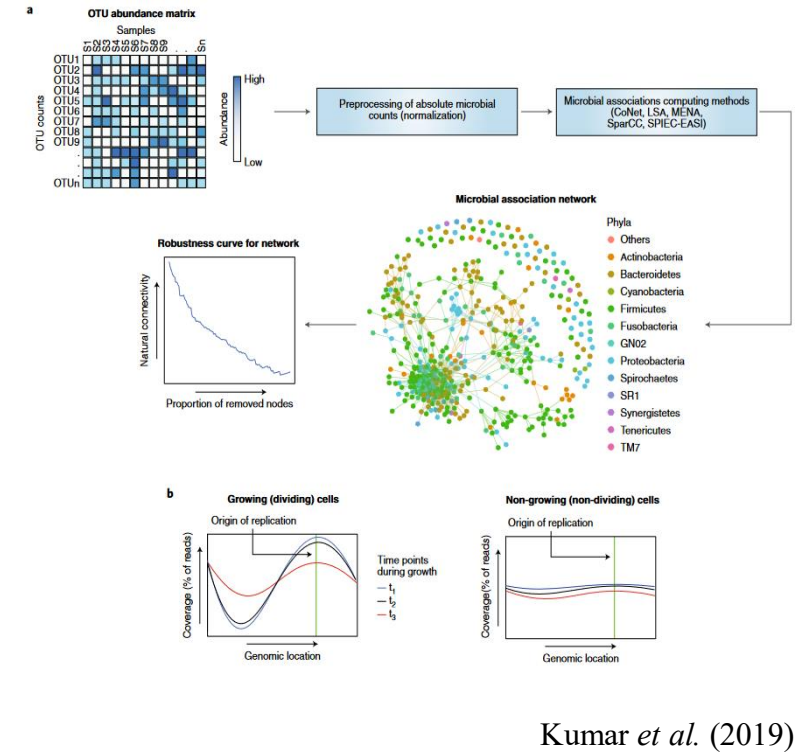
- Marine microbial community constitute 70% of the living biomass in the global ocean and they play pivotal roles in ocean ecosystem food webs and global biogeochemical cycles.

# Marine microbial community interactions

## Microbial interaction



## High throughput sequencing

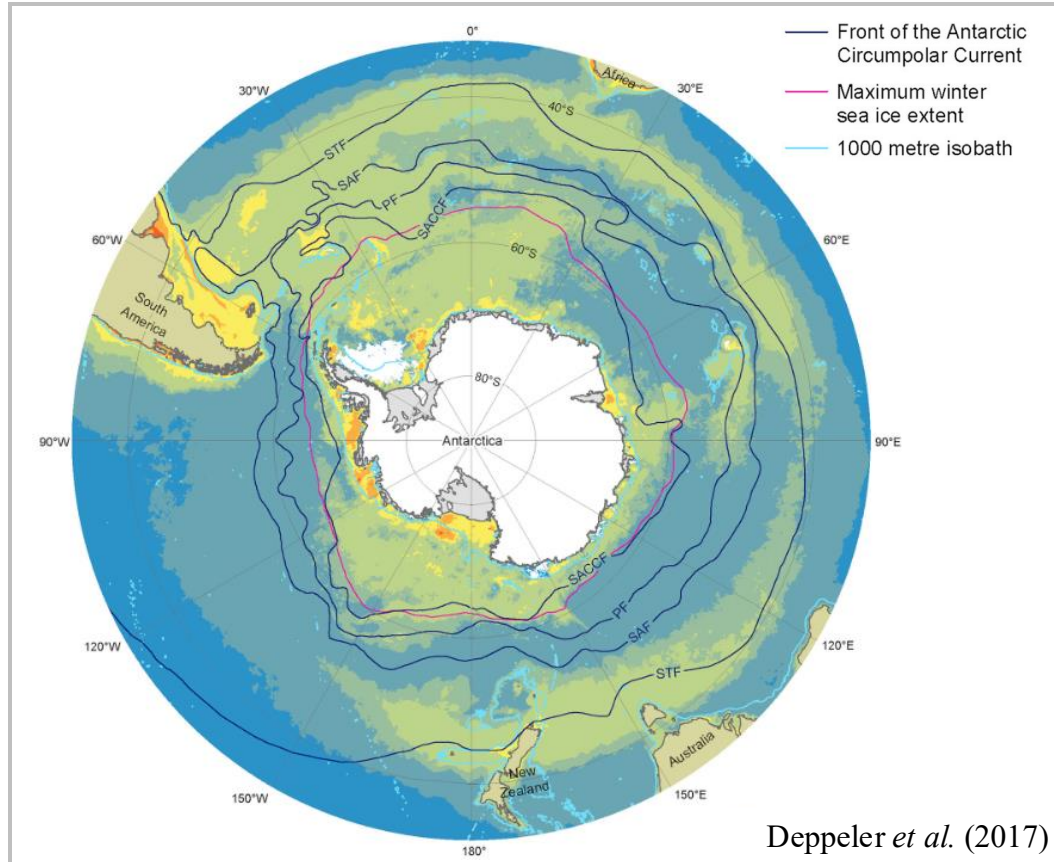


- Structuring marine microbial community composition are selection by biotic interaction.
- eDNA sequencing able to address the large complexity of microbial interactions.

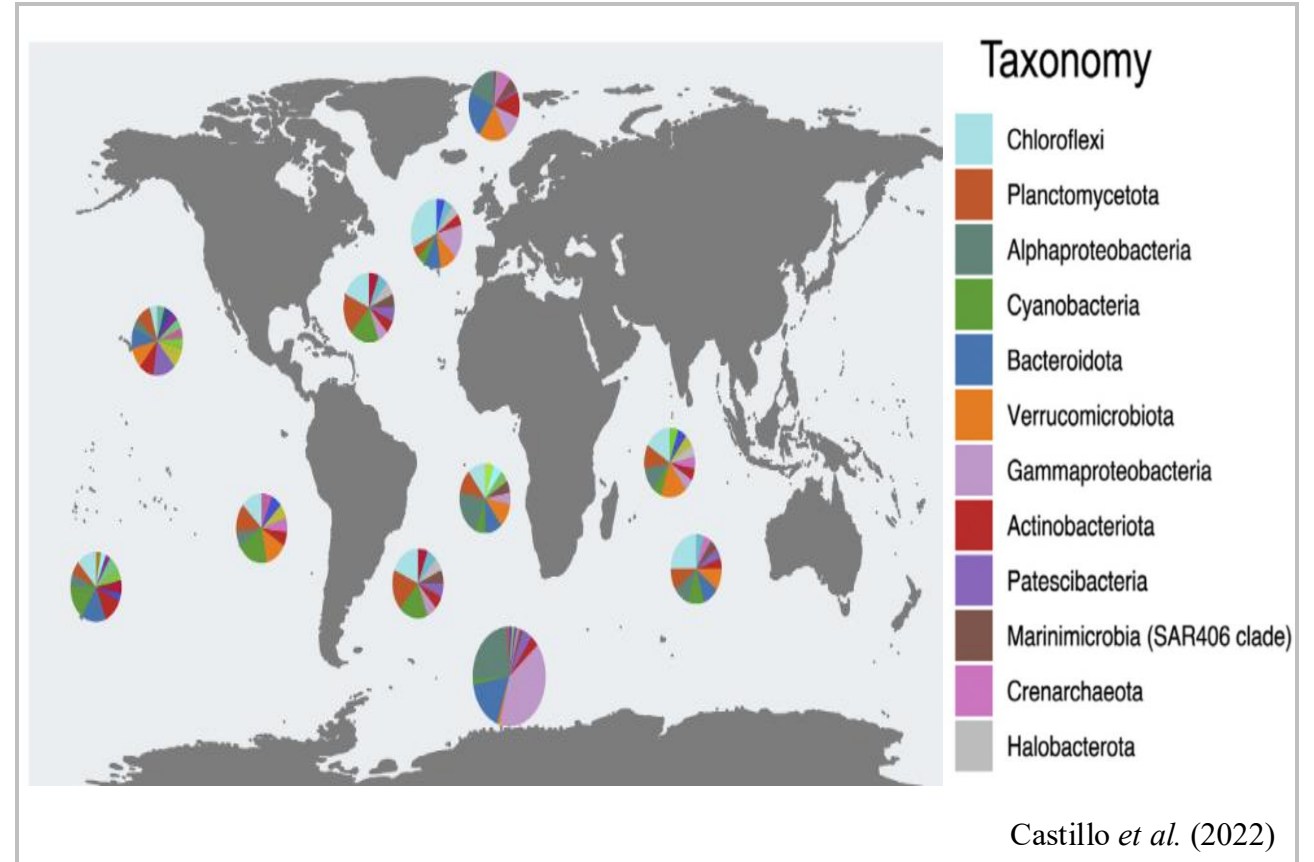


# Southern Ocean

## Southern Ocean environment



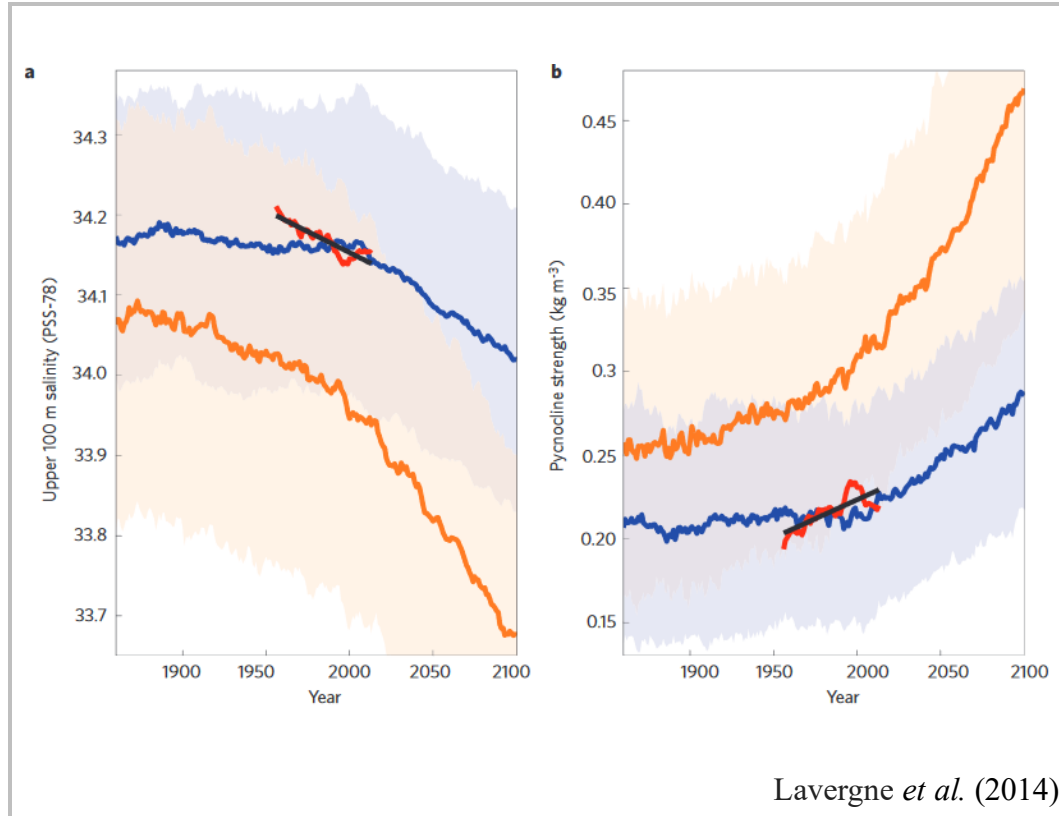
## Unique microbial community structure



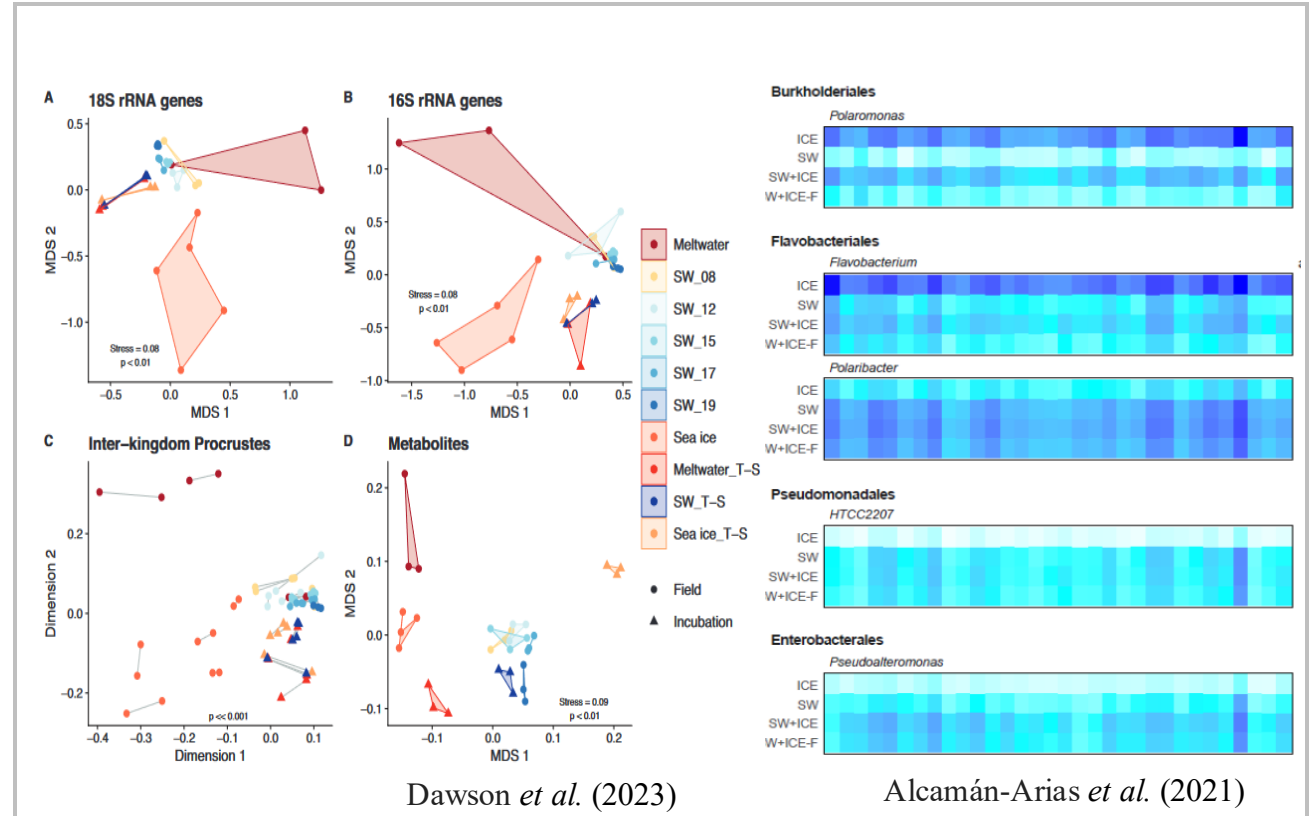
- The Southern Ocean accounts for approximately 25% of the oceanic uptake of atmospheric CO<sub>2</sub> despite only representing 20% of the total ocean surface area (*Castillo et al. 2022*).

# Southern Ocean freshening

## Freshening in Southern Ocean

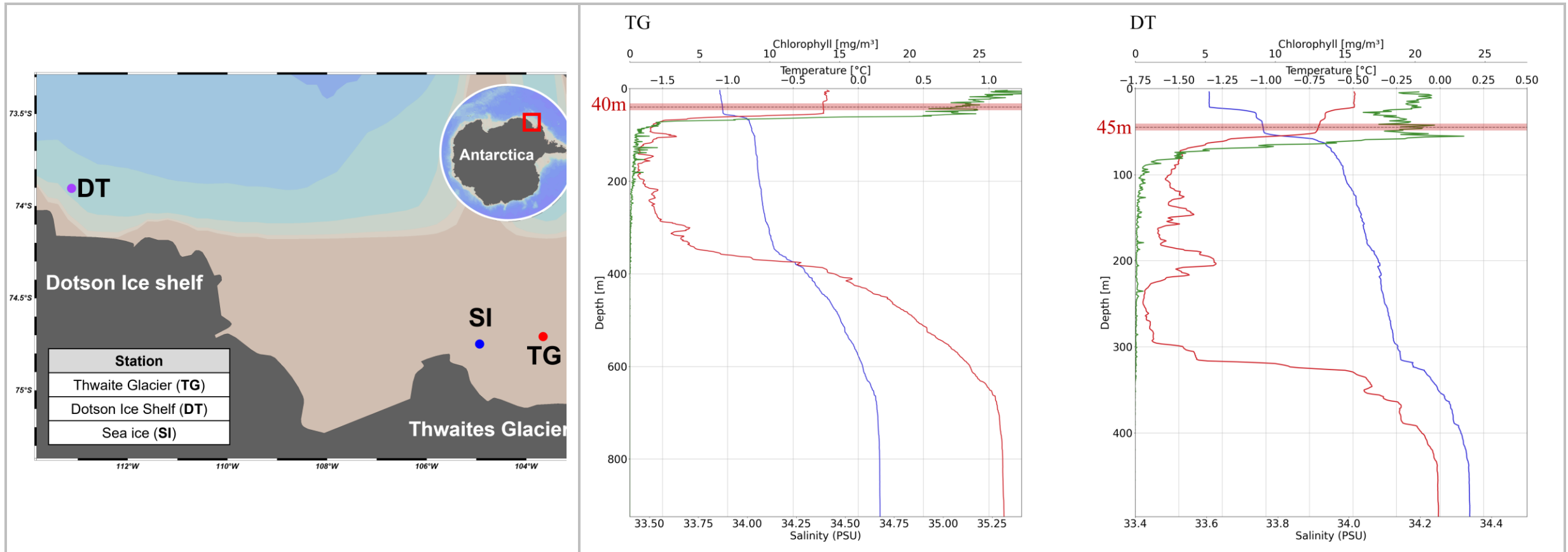


## Freshening microcosm experiment



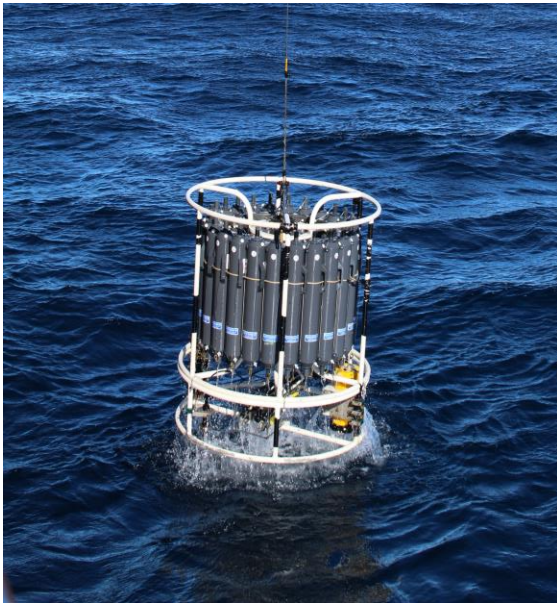
- Climate change-driven surface freshening in the Southern Ocean is expected to intensify salinity stratification, impacting microbial community structure, interactions, and functions.

## Materials and methods – sampling

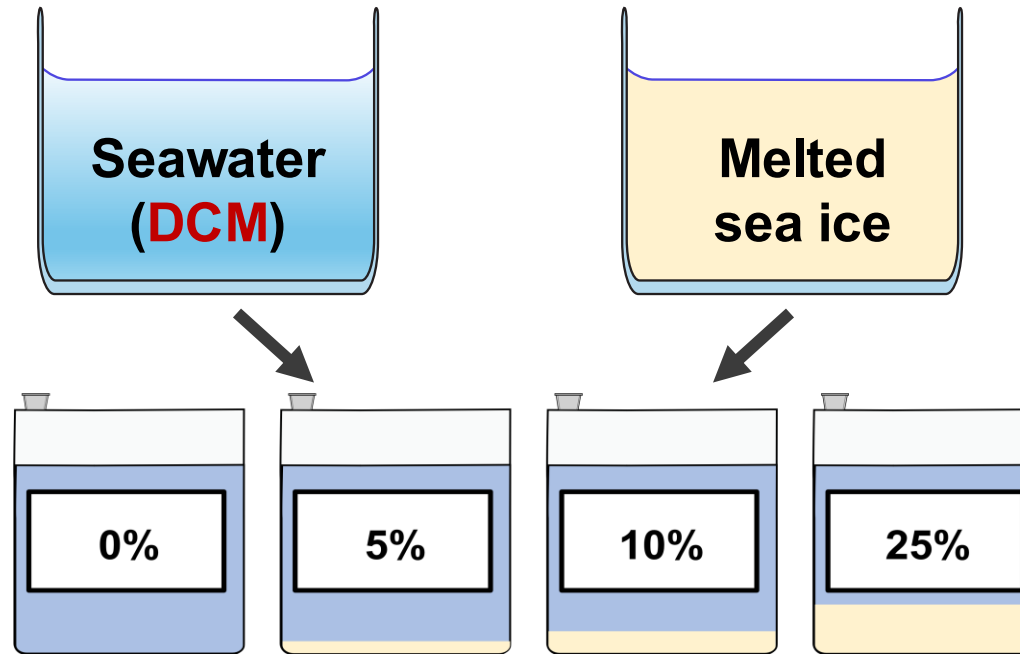


- CTD with Niskin bottle were used for seawater sample during **January 7<sup>th</sup> – 31<sup>st</sup>, 2024**.
- **Deep chlorophyll maximum** (DCM) layer was sampled for microcosm experiment.

## Materials and methods – experimental procedure



| Sampled depth |
|---------------|
| TG DCM: 40 m  |
| DT DCM: 45 m  |



| Incubation temperature (°C) |
|-----------------------------|
| TG samples : -1             |
| DT samples : 1              |

| Subsampling |
|-------------|
| Day 2 (T2)  |
| Day 7 (T7)  |



| Salinity (PSU) |
|----------------|
| TG DCM: 33.8   |
| DT DCM: 34.0   |
| Sea ice : 3.3  |

- Niskin bottle were used for seawater sample at DCM layer during 2023-2024 Antarctic expedition.
- Dilution microcosm experiment were conducted in *in situ* temperature with light ( $40 \mu\text{mol m}^{-2} \text{s}^{-1}$ ).

## Materials and methods – experimental procedure

### Analysis

#### 20 µm filtration

- Microphytoplankton (>20 µm)
- Particle-attached (PA) prokaryotes

#### 3 µm filtration

- Nanophytoplankton (3 – 20 µm)
- Particle-attached (PA) prokaryotes

#### 0.2 µm filtration

- Picophytoplankton (0.2 – 3 µm)
- Free-living (FL) prokaryotes

### Bacterial abundance

### Metabolic activity

#### Filtration

0.5 L of each subsample (triplicate)

#### DNA extraction & PCR

16S/18S rRNA gene (515Y/926R primer set)

Parada *et al.* (2019)

#### Sequencing (NGS)

Illumina MiSeq platform PE300

#### Sequence processing

FastQC, Trimmomatic, Cutadapt, Qiime2, R

#### Bacterial abundance

Flow cytometry (SYBR green)

#### Community activity

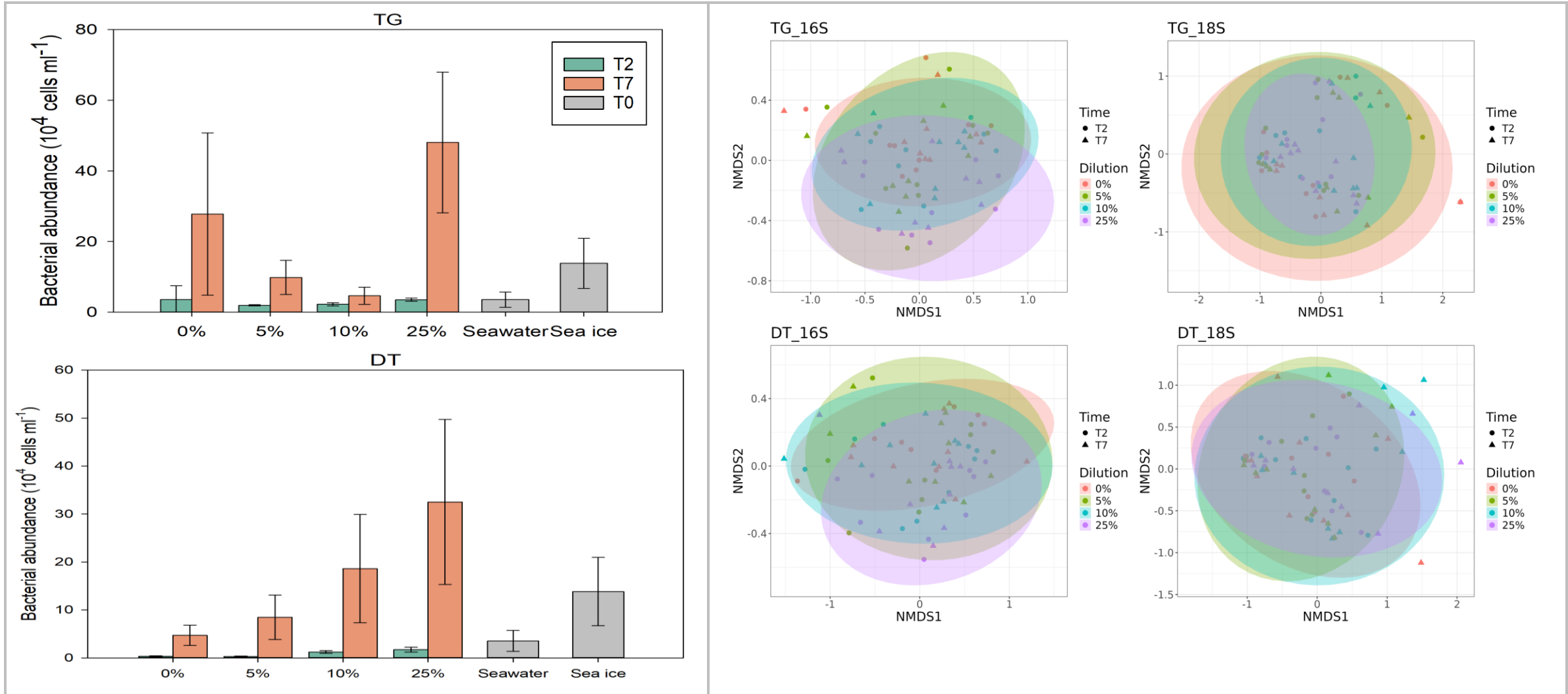
RT-qPCR, qPCR (16S, 18S rRNA/rDNA ratio)

## Data analysis

- Community **diversity**
- Bacterial **abundance**
- **Taxonomic** analysis
- **Co-occurrence** network analysis (FlashWeave)
- rDNA, rRNA ratio analysis

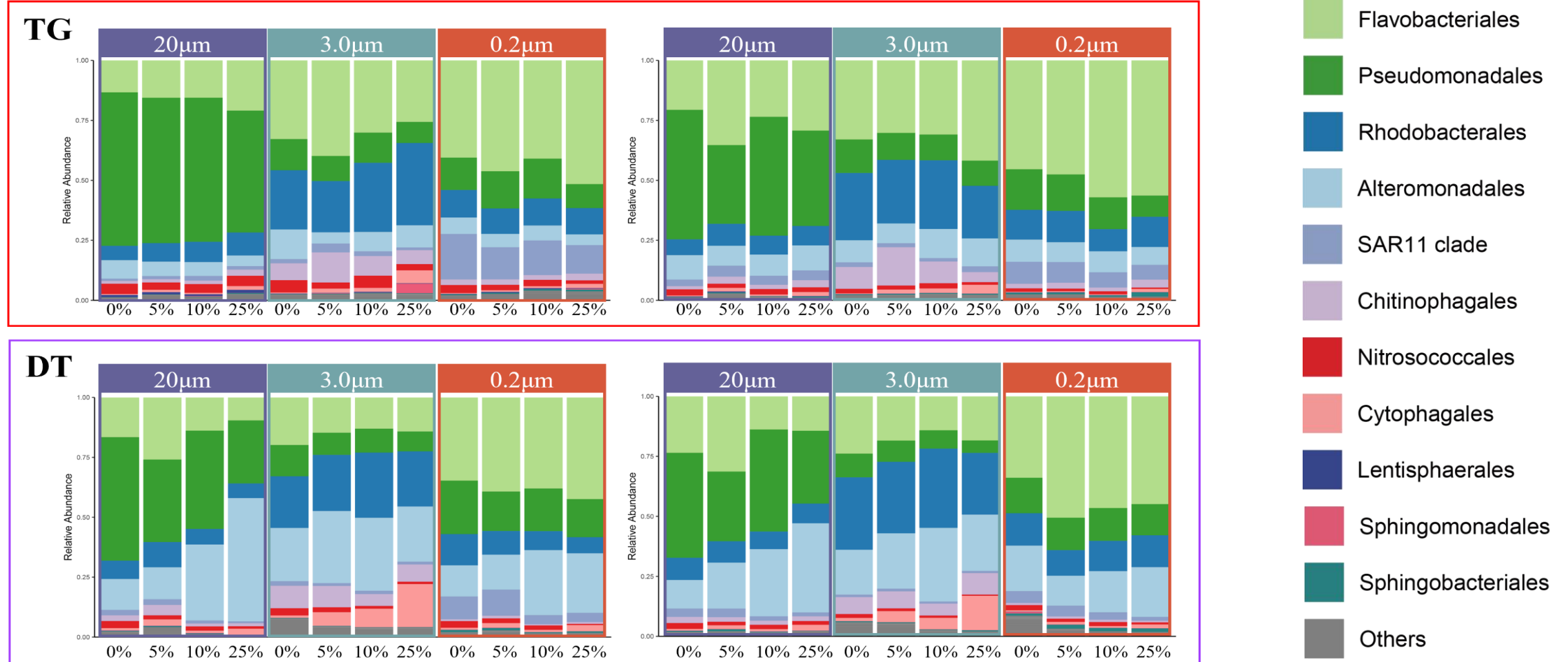


## Diversity of microbial community



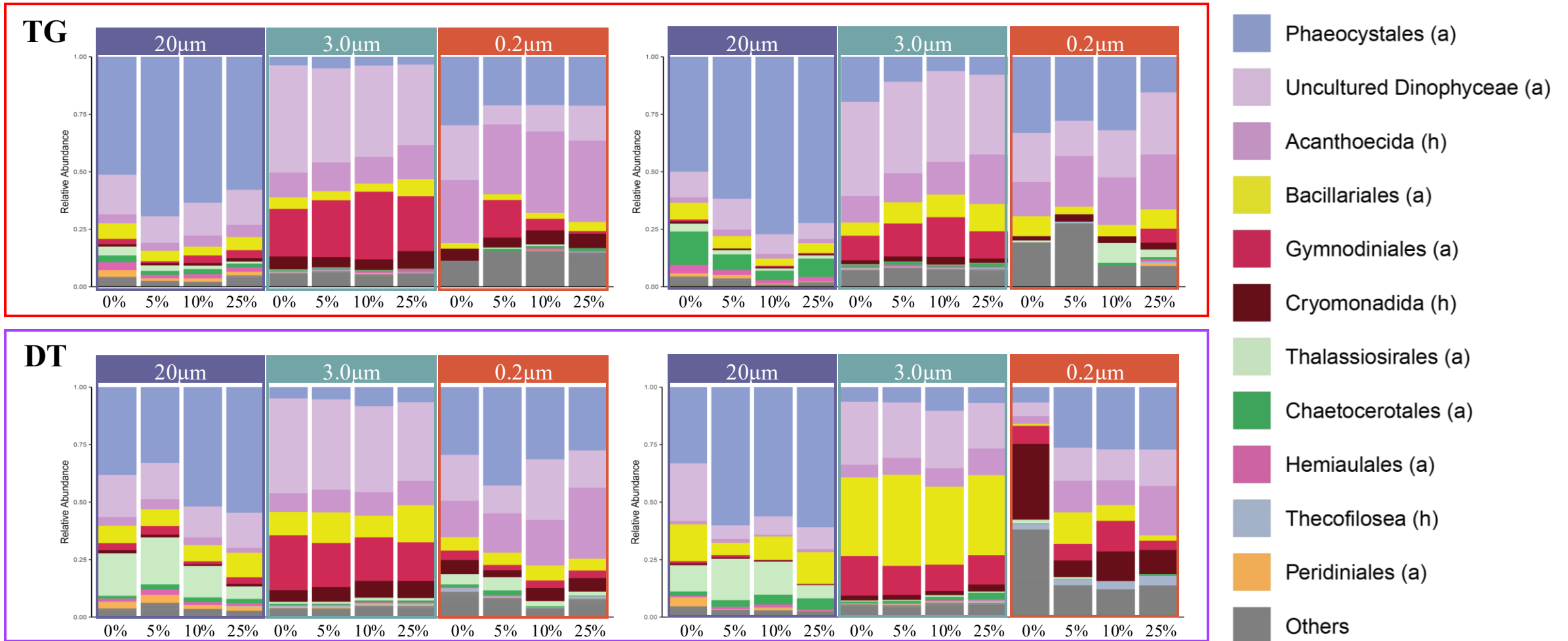
- Both prokaryotic, eukaryotic community is a stronger influence by the **dilution level** on the overall composition than the time interval measured.

## Community after incubation



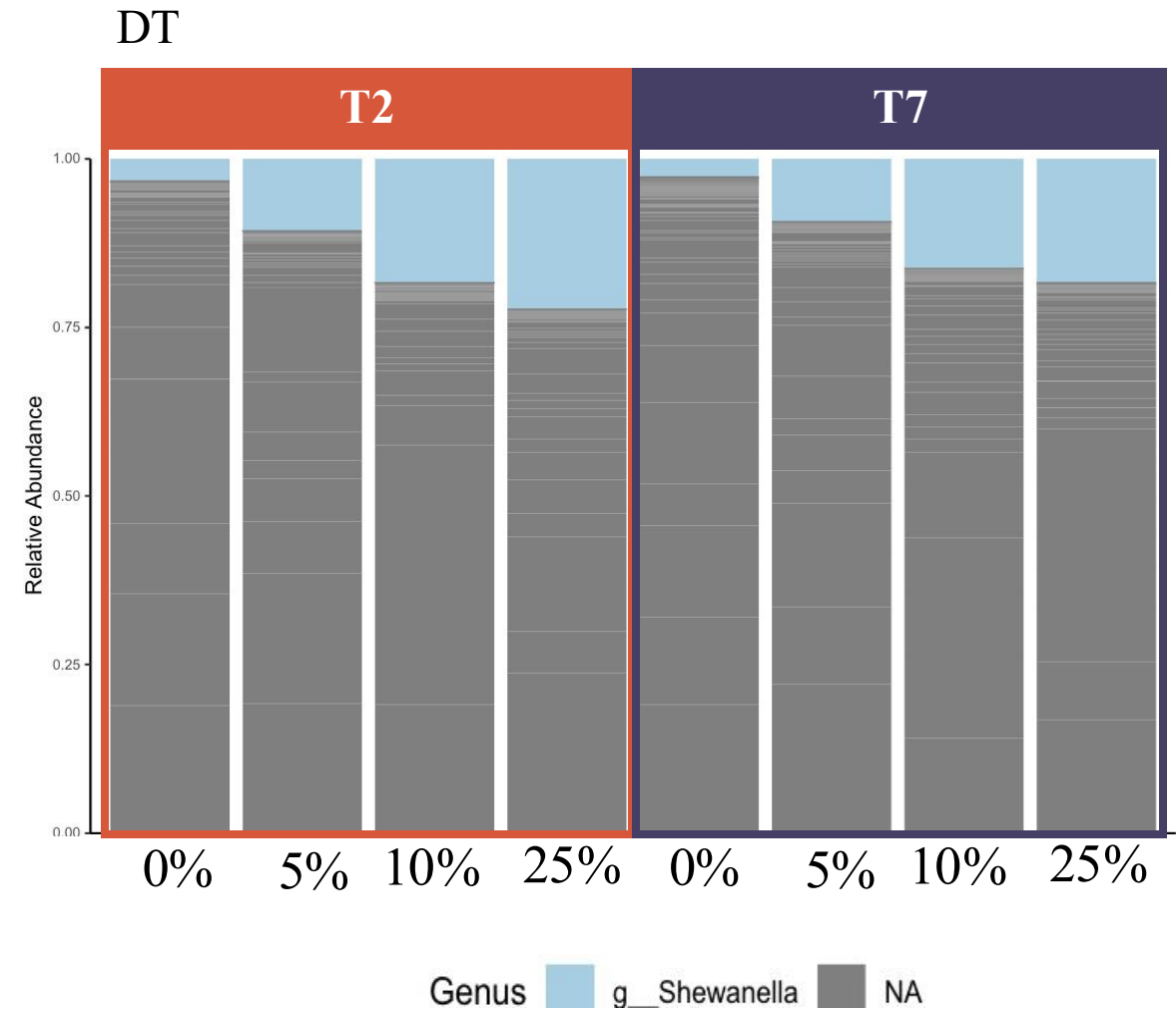
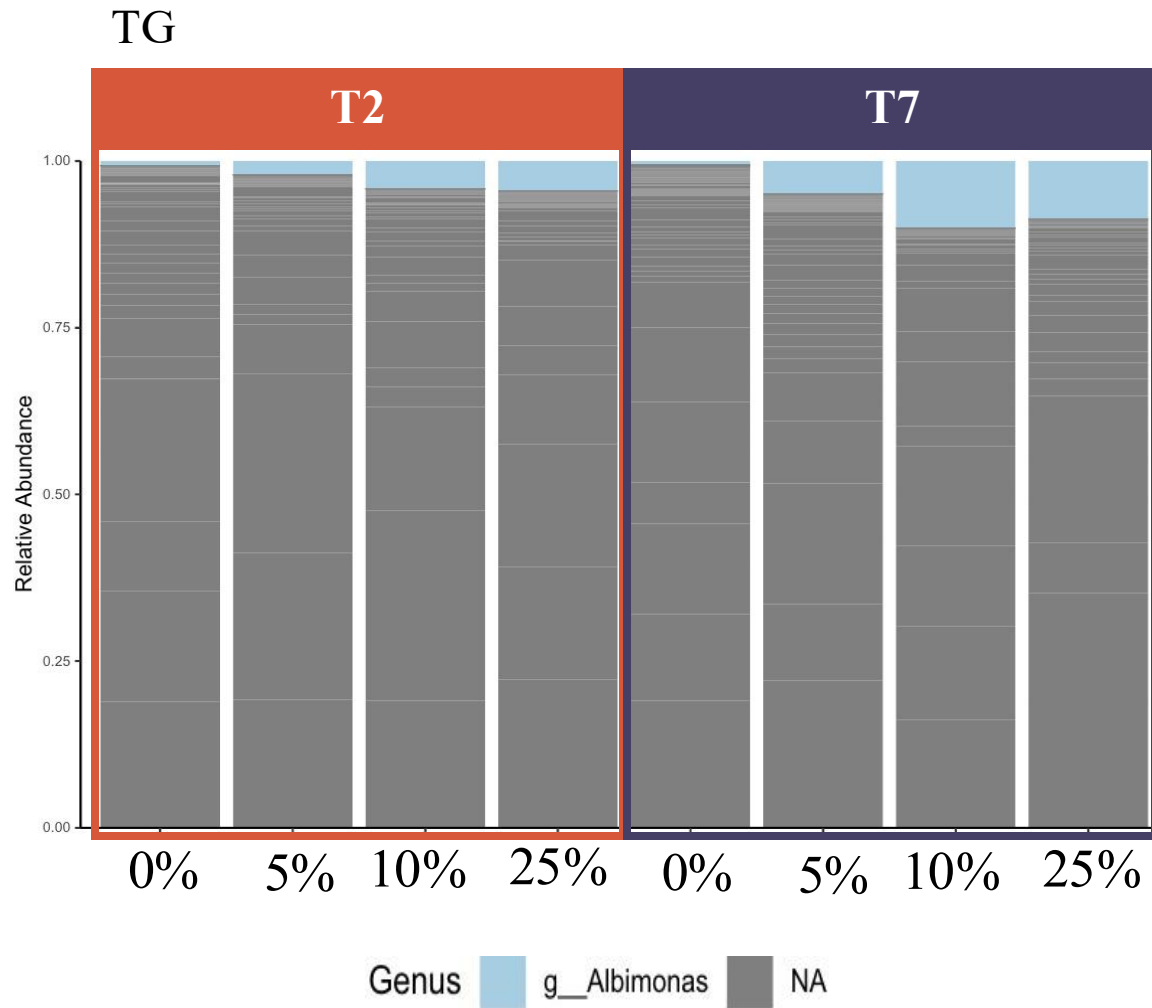
- Flavobacteriales significantly increased as the dilution increased for TG.
- In DT region, Alteromonadales and Cytophagales increased as the dilution increases.

## Community after incubation



- Phaeocystales, Bacillariales, Gymnodiniales increase as the dilution increased in TG.
- Phaeocystales increased while Thalassiosirales decreased with higher dilution in DT.

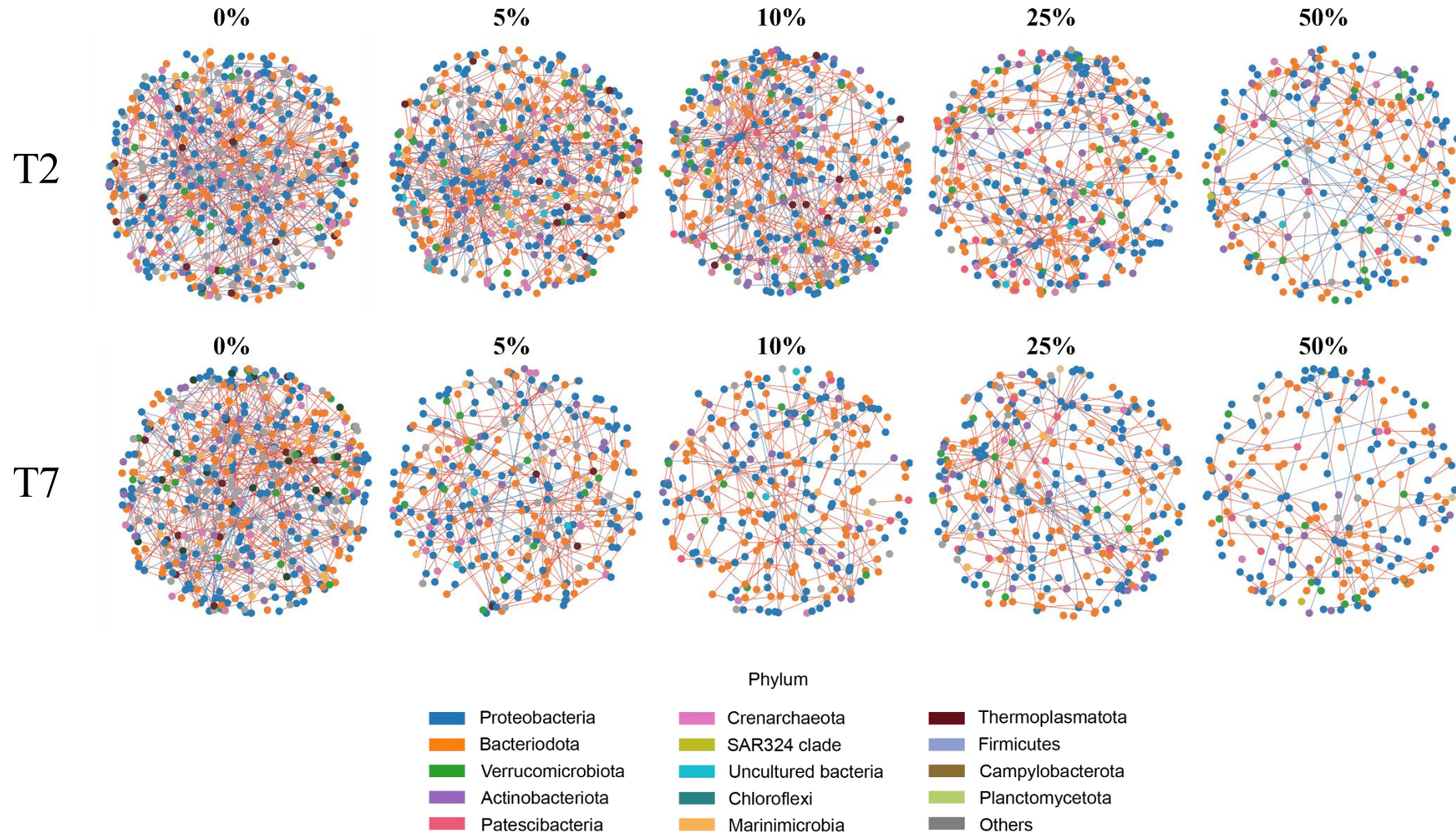
## Community after incubation



- The abundance of *Albimonas* sp. and *Shewanella* sp. exhibited an increased with the freshening.
- May serve as biological sentinels for monitoring the freshening in Southern Ocean.

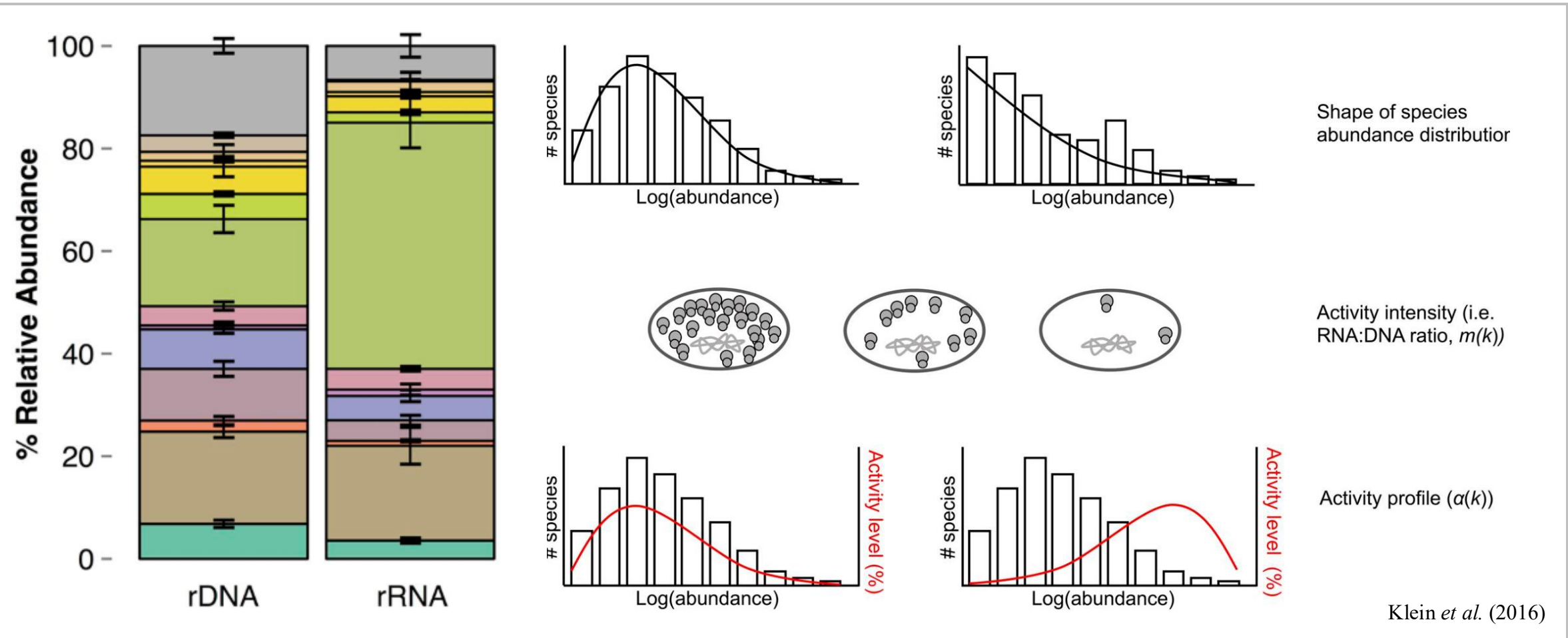


## Co-occurrence network



- Co-occurrence network analysis showed that increased dilution weakened prokaryotic community interactions by reducing connection at higher meltwater concentrations.

# Metabolic activity of microbial community



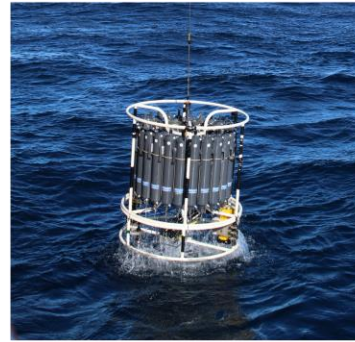
- rRNA/rDNA ratio indicates greater metabolic activity, is an effective method for assessing microbial responses to environmental changes and will be used to evaluate phytoplankton and prokaryote activity

## Conclusion

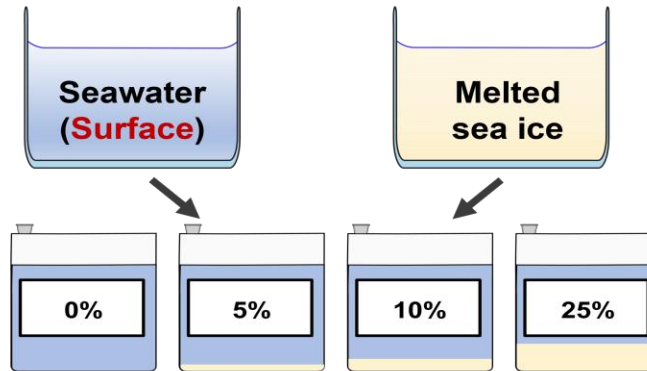
- eDNA sequencing of 16S, and 18S rDNA sequencing confirmed that microbial communities are sensitive to meltwater intrusion simulation at Southern Ocean.
- Observed distinct, site-specific changes in community composition, demonstrating significant spatial variability in how marine ecosystems will respond to freshening.
- Some genera may potentially serve as biological sentinels for monitoring the freshening of the Southern Ocean.
- Increased freshening significantly destabilized microbial networks, directly impacting community interactions and potentially compromising critical ecosystem functions in the Southern Ocean.



## Future study



| Sampled depth |
|---------------|
| TG DCM: 40 m  |
| DT DCM: 45 m  |



| Incubation temperature (°C) |
|-----------------------------|
| TG samples : -1             |
| DT samples : 1              |

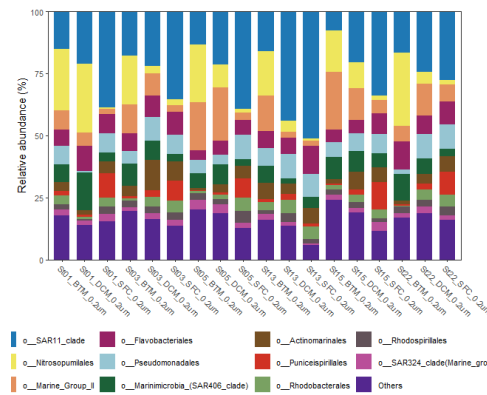
| Subsampling |
|-------------|
| Day 2 (T2)  |
| Day 7 (T7)  |



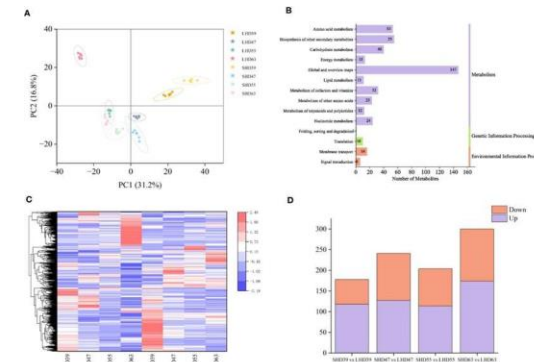
| Salinity (PSU) |
|----------------|
| TG DCM: 33.8   |
| DT DCM: 34.0   |
| Sea ice : 3.3  |



## Metagenomics



## Metabolomics



- Microbial interactions on both metagenomics and metabolomics to reveal the community and the metabolite changes in the Bering strait.



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**Thank you**