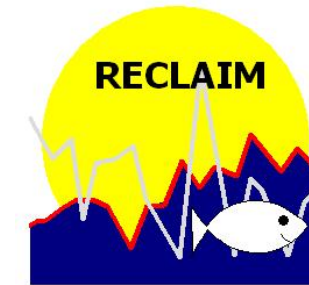


Larval Fish Physiology & Individual-based Models: Exploring Climate Impacts on Early Life Stages of Key Species

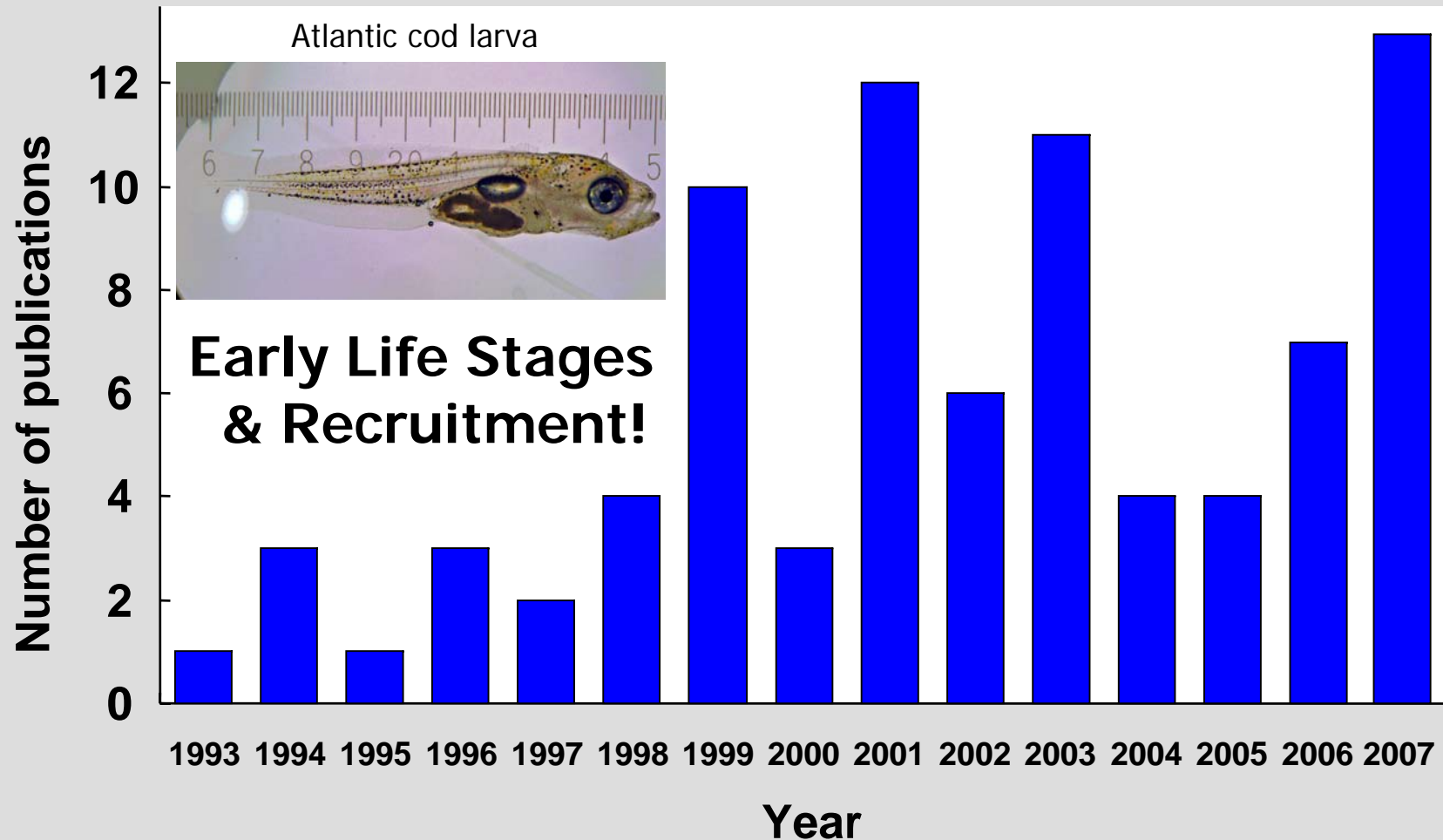
Myron A. Peck¹, Ute Daewel¹ & Corinna Schrum²

¹Center for Marine and Climate Research
Institute of Hydrobiology and Fisheries Science
University of Hamburg
Hamburg, Germany

²Geophysical Institute, University of Bergen
Bjerknes Centre for Climate Research
Bergen, Norway



Coupled physical – biological IBMs for marine fish larvae



IBMs allow individuals to have unique characteristics – in the case of pelagic eggs & larvae - environmental conditions in time & space

(updated from Tom Miller 2006, WKAMF)

coupled physical-biological model system

egg & larval IBM parameterization

Part I

endogenous feeding
stages

development of eggs &
yolksac larvae

Temperature

Part II

exogenously feeding
stages

rates of energy loss &
gain by larvae

Temperature

Part III model application (North Sea)



1-D IBM to 3-D coupled models



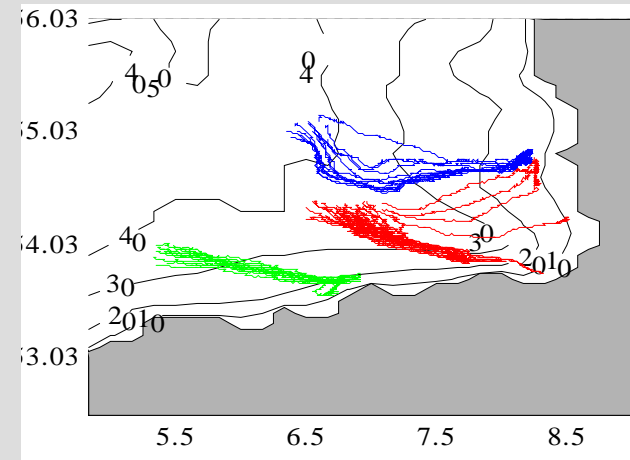
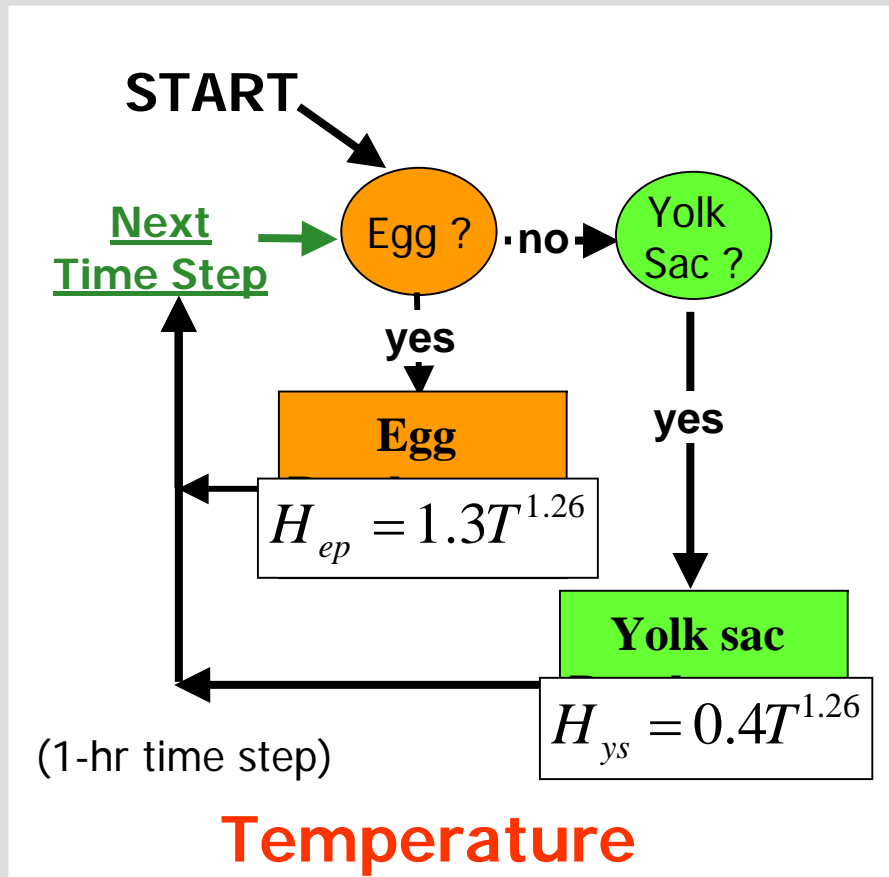
demersal gadid vs. pelagic clupeid

(*Gadus morhua*)

(*sprattus sprattus*)

larval vital rates, temperature & prey availability

Part I: Eggs and Yolksac Larvae (simple IBM's)



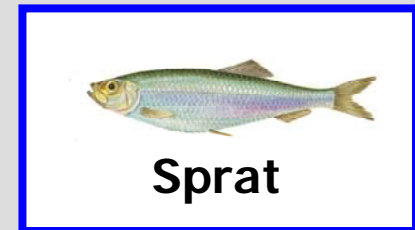
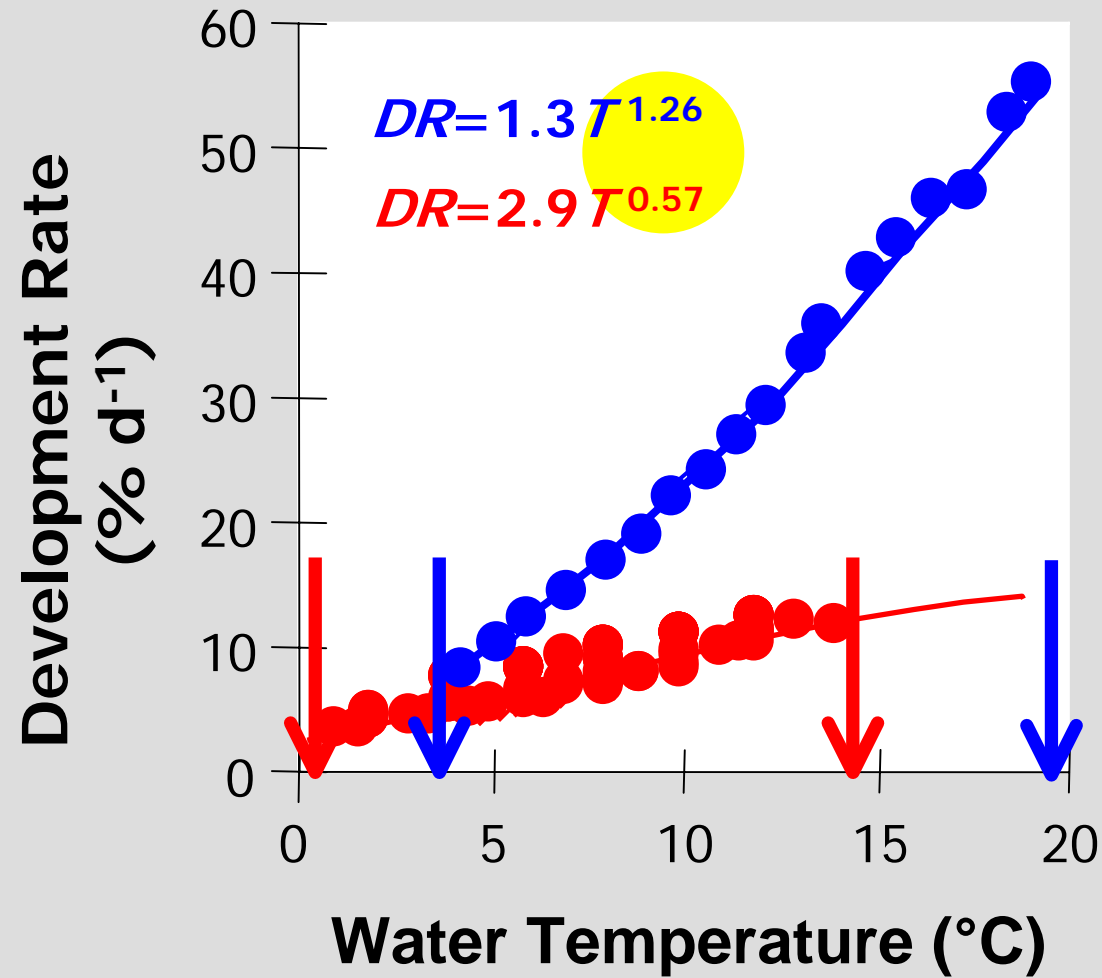
drift simulations

Climate impacts on:

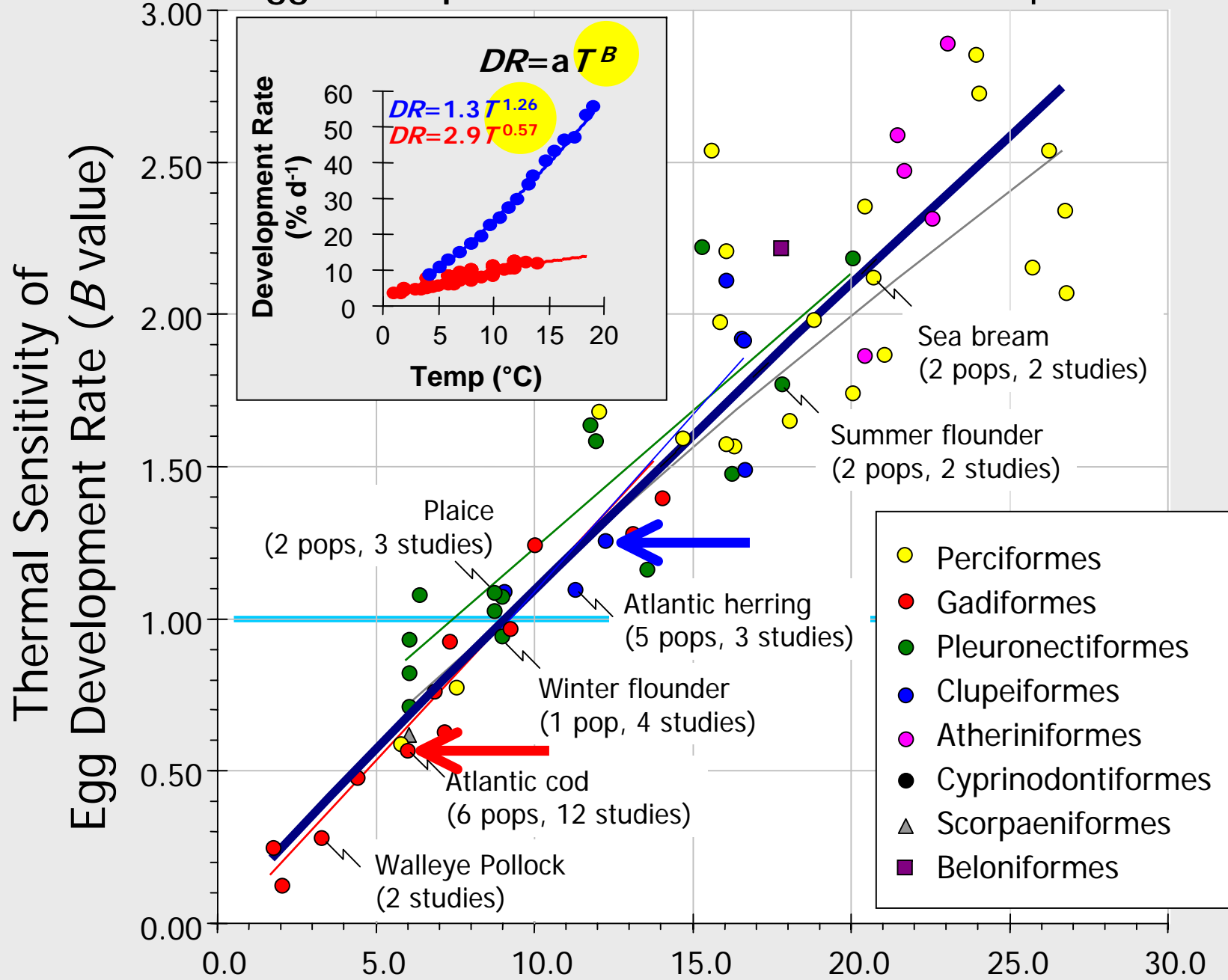
- Transport
- Habitat connectivity
- Life cycle closure

Egg Development Rates vs. Temperature

$$DR = a T^B$$



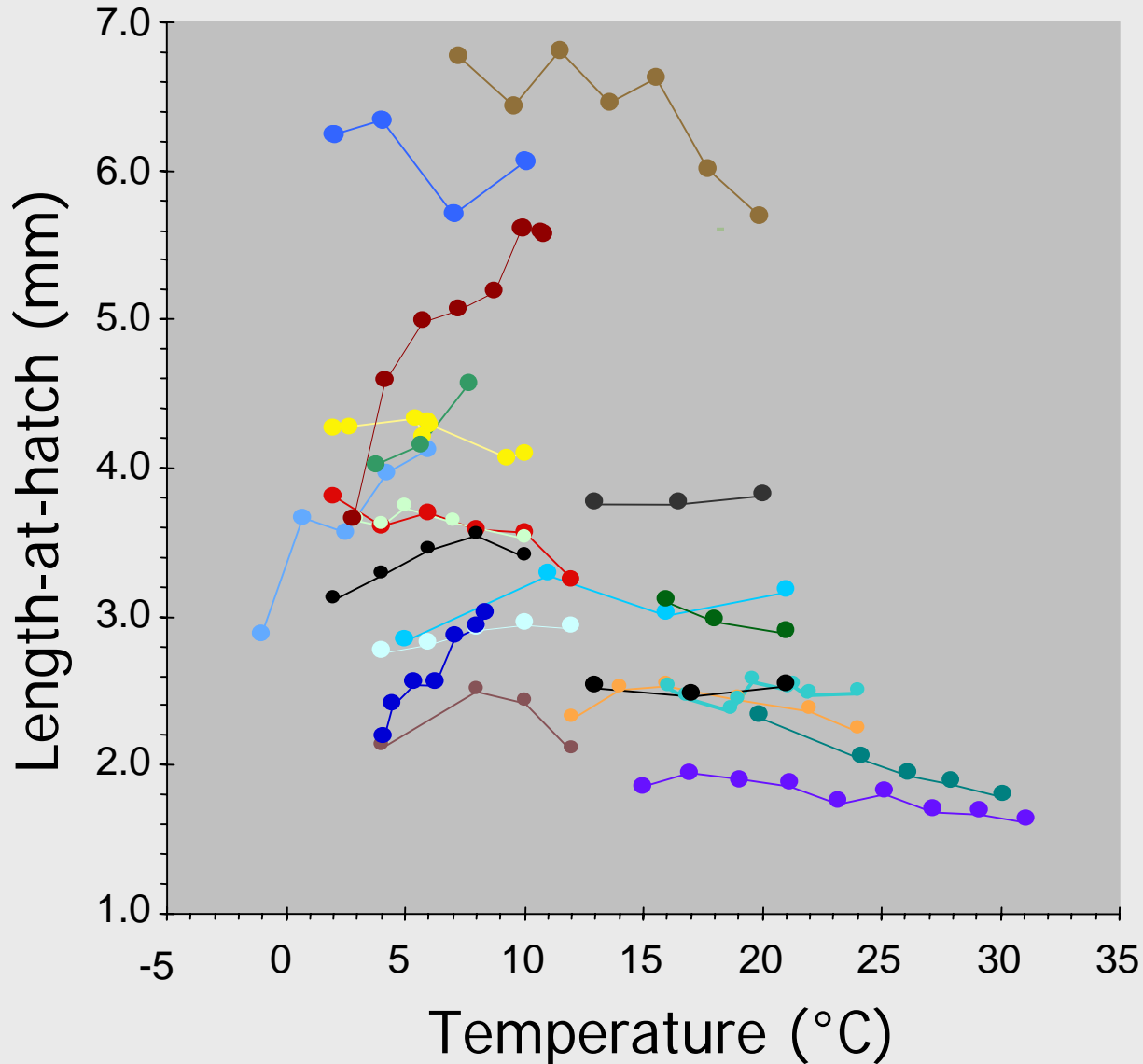
Egg Development Rates for 64 Marine Fish Species



(Peck, Chambers, Geffen, data compilation)

Mean Incubation Temperature (°C)

Yolksac Larvae: Length-at-hatch (22 marine species)

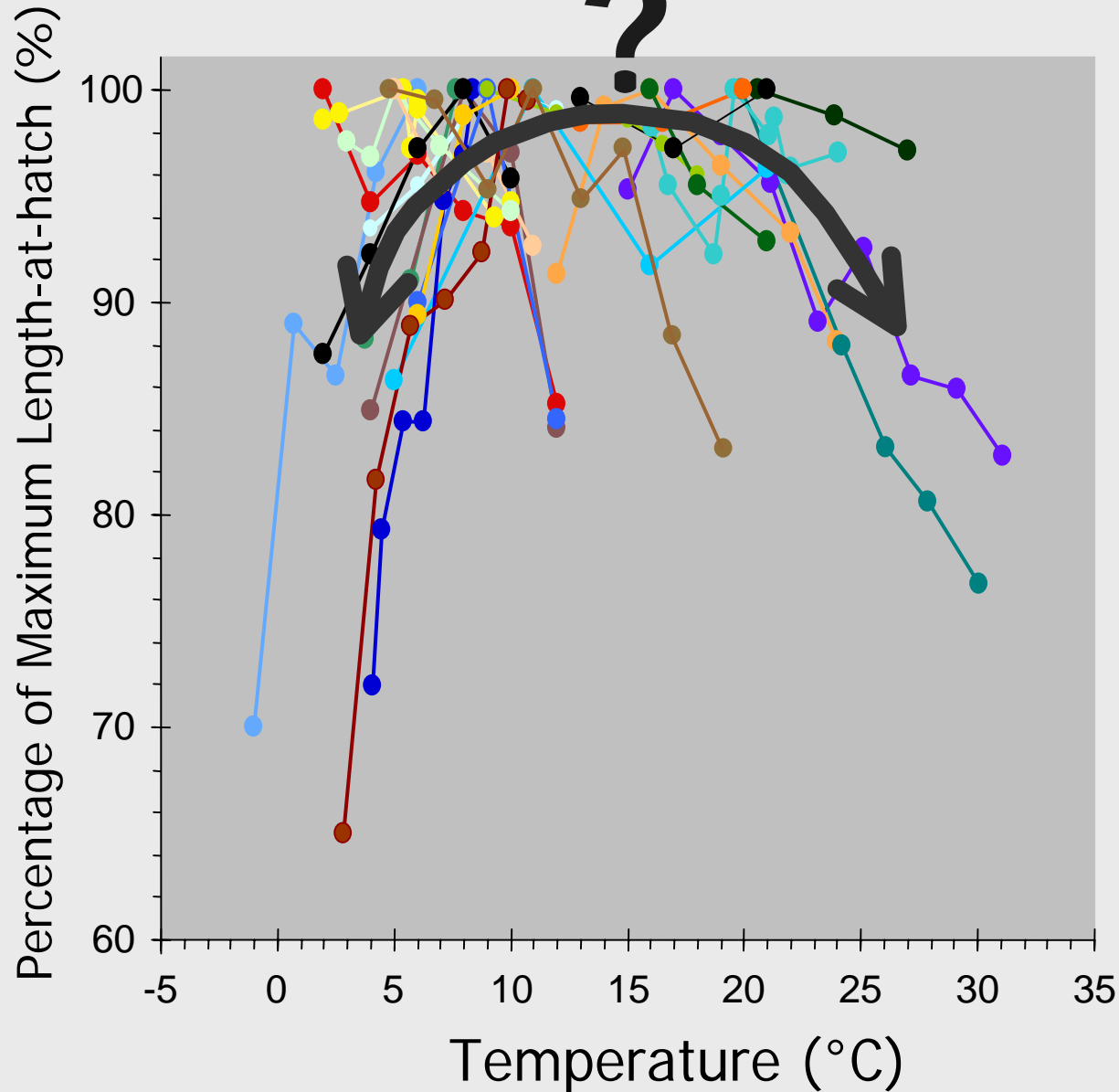


- *Anarhichas lupus*
- *Bairdiella icistia*
- *Clupea harengus*
- *Clupea harengus*
- *Eopsetta jordani*
- *Engraulis encrasicolus*
- *Gadus morhua*
- *Gadus morhua*
- *Gadus macrocephalus*
- *Hippoglossoides elassodon*
- *Limanda ferruginea*
- *Morone americana*
- *Myteroperca rosacea*
- *Melanogrammus aeglefinus*
- *Ophiodon elongatus*
- *Pagellus erythrinus*
- *Rhombosolea tapirina*
- *Pagrus major*
- *Paralichthys dentatus*
- *Parophrys vetulus*
- *Pseudopleuronectes americanus*
- *Sardinops sagax musica*
- *Sparus aurata*
- *Theragra chalcogramma*

(M. Peck, & F. Bils unpublished compilation)



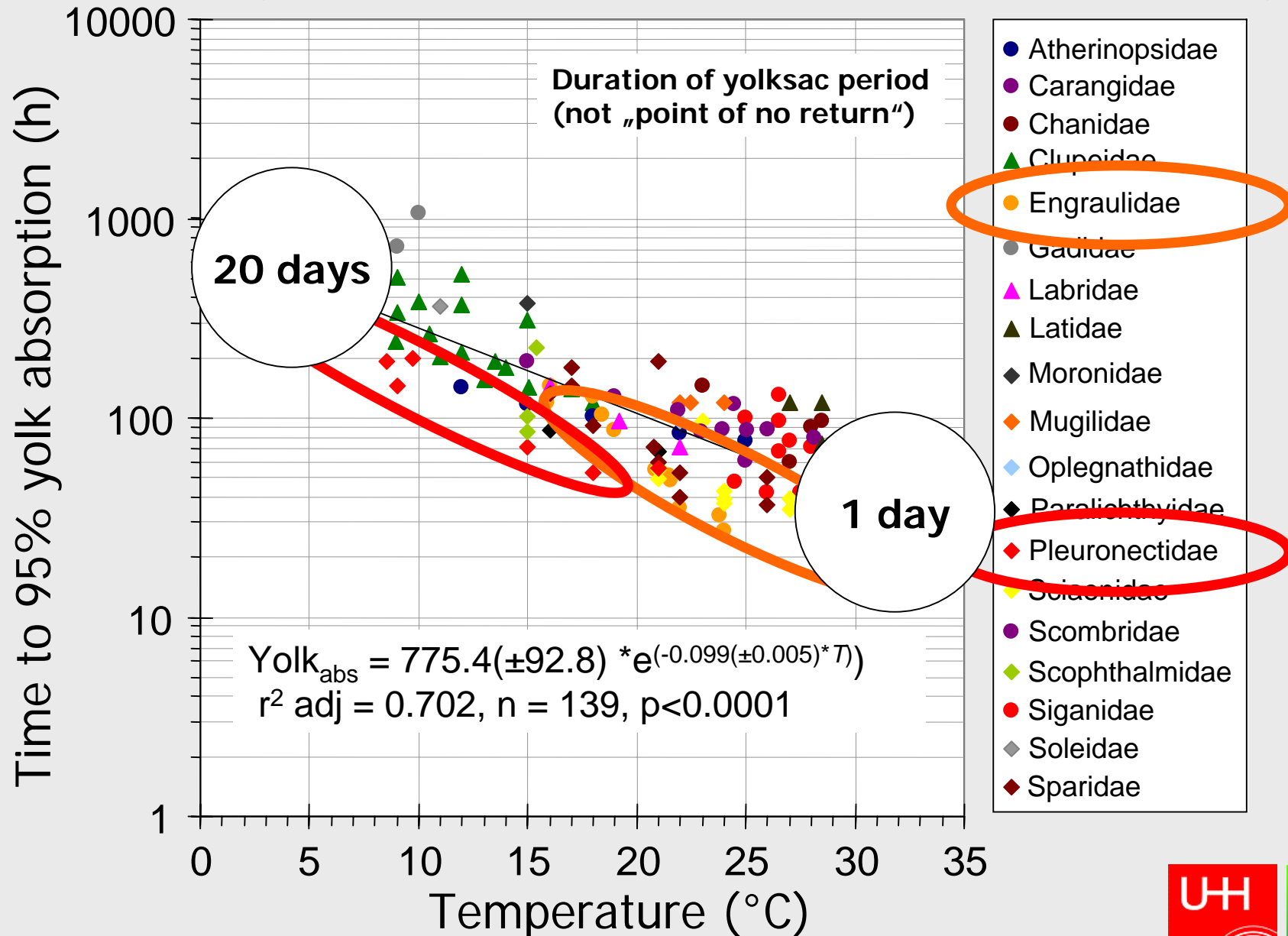
Normalized to Max Length-at-hatch



- Anarhichas lupus*
- Bairdiella icistia*
- Clupea harengus*
- Eopsetta jordani*
- Engraulis encrasicolus*
- Gadus morhua*
- Gadus macrocephalus*
- Hippoglossoides elassodon*
- Limanda ferruginea*
- Morone americana*
- Myteroperca rosacea*
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- Sardinops sagax musica*
- Sparus aurata*
- Theragra chalcogramma*



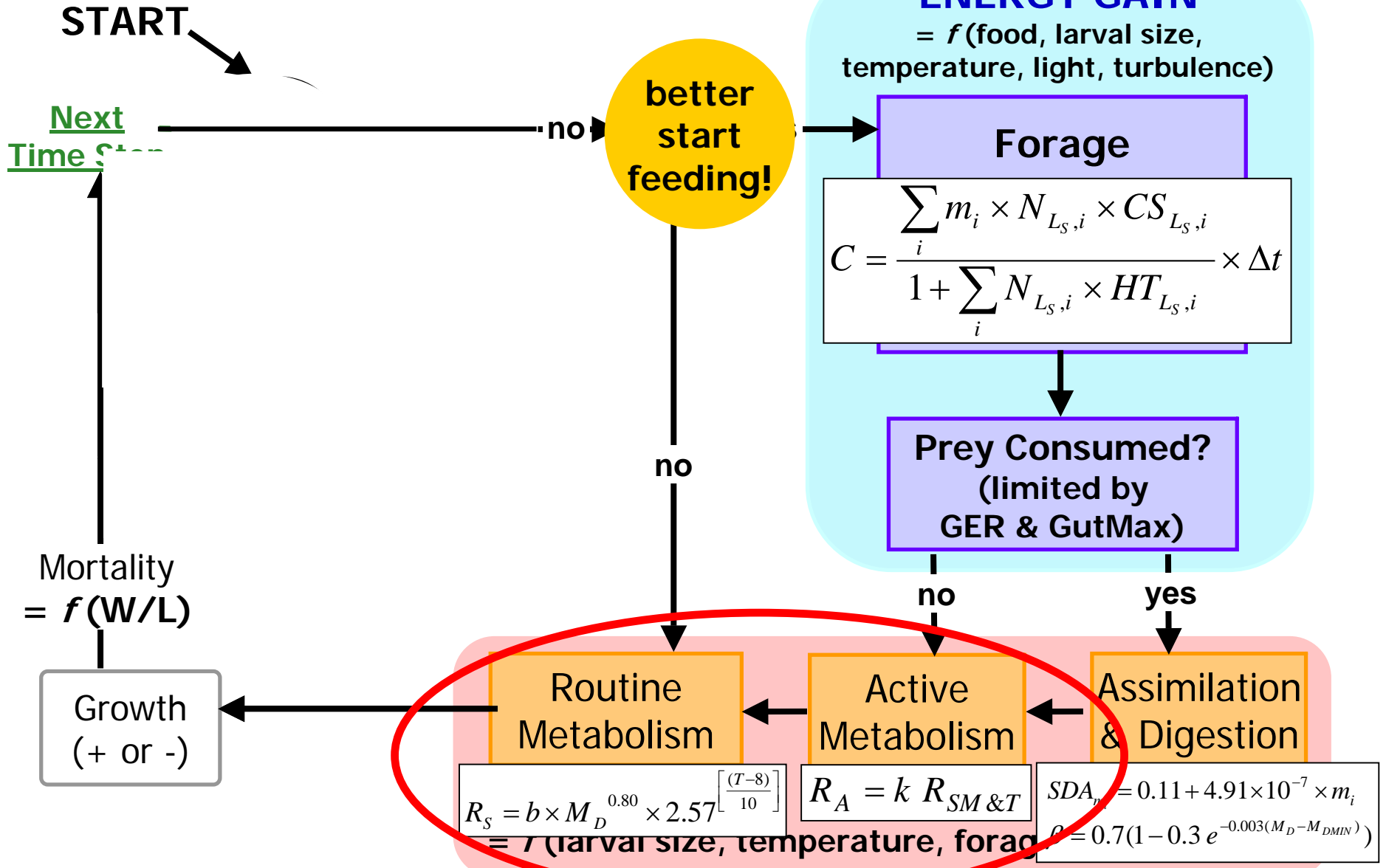
Best (but difficult) comparison is of individual slopes (more negative slope = most impacted by warming)



(M. Peck, & C. Lindemann unpublished compilation)

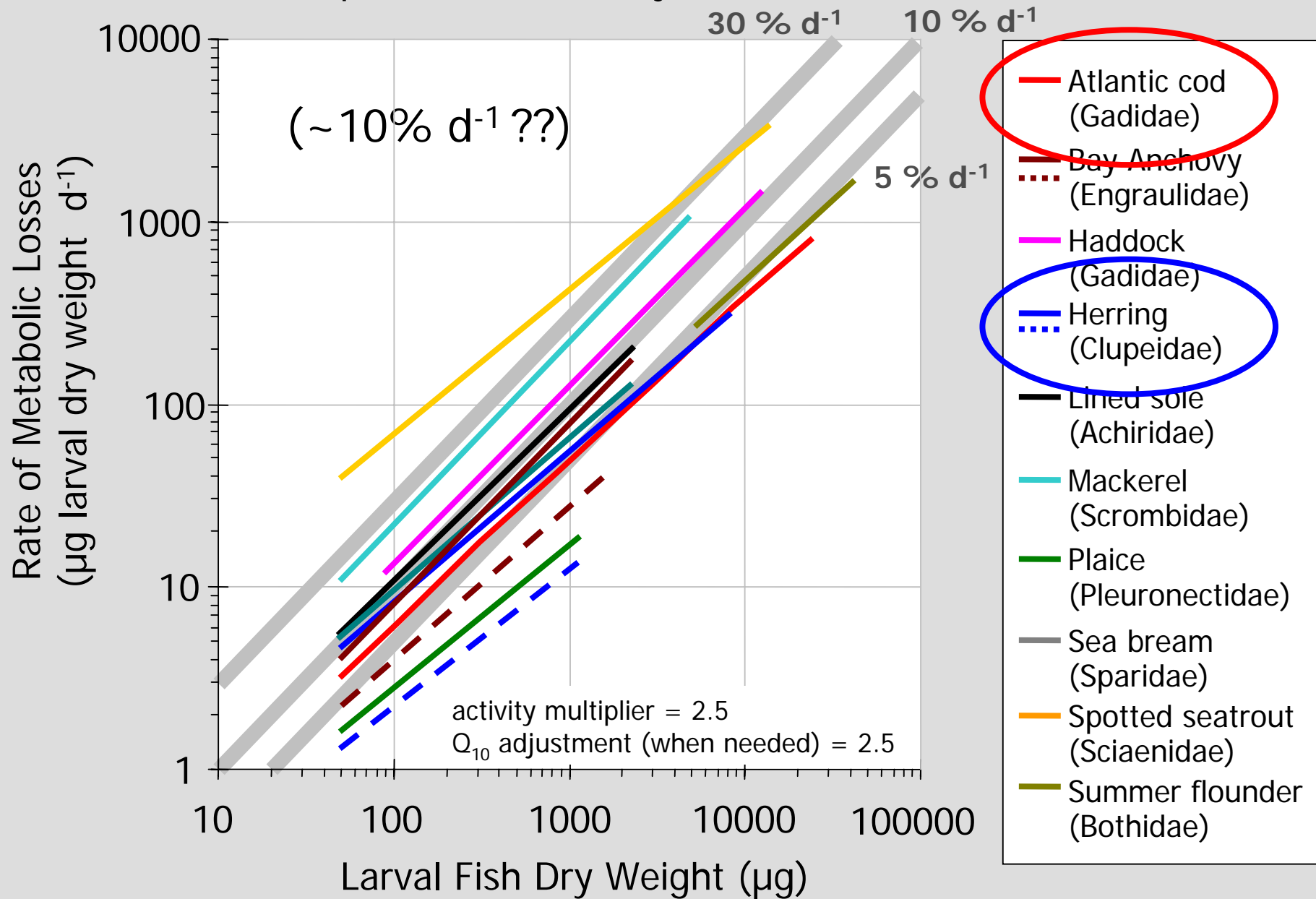


**Part II: Exogenously-feeding larvae
Foraging and Growth Subroutines**

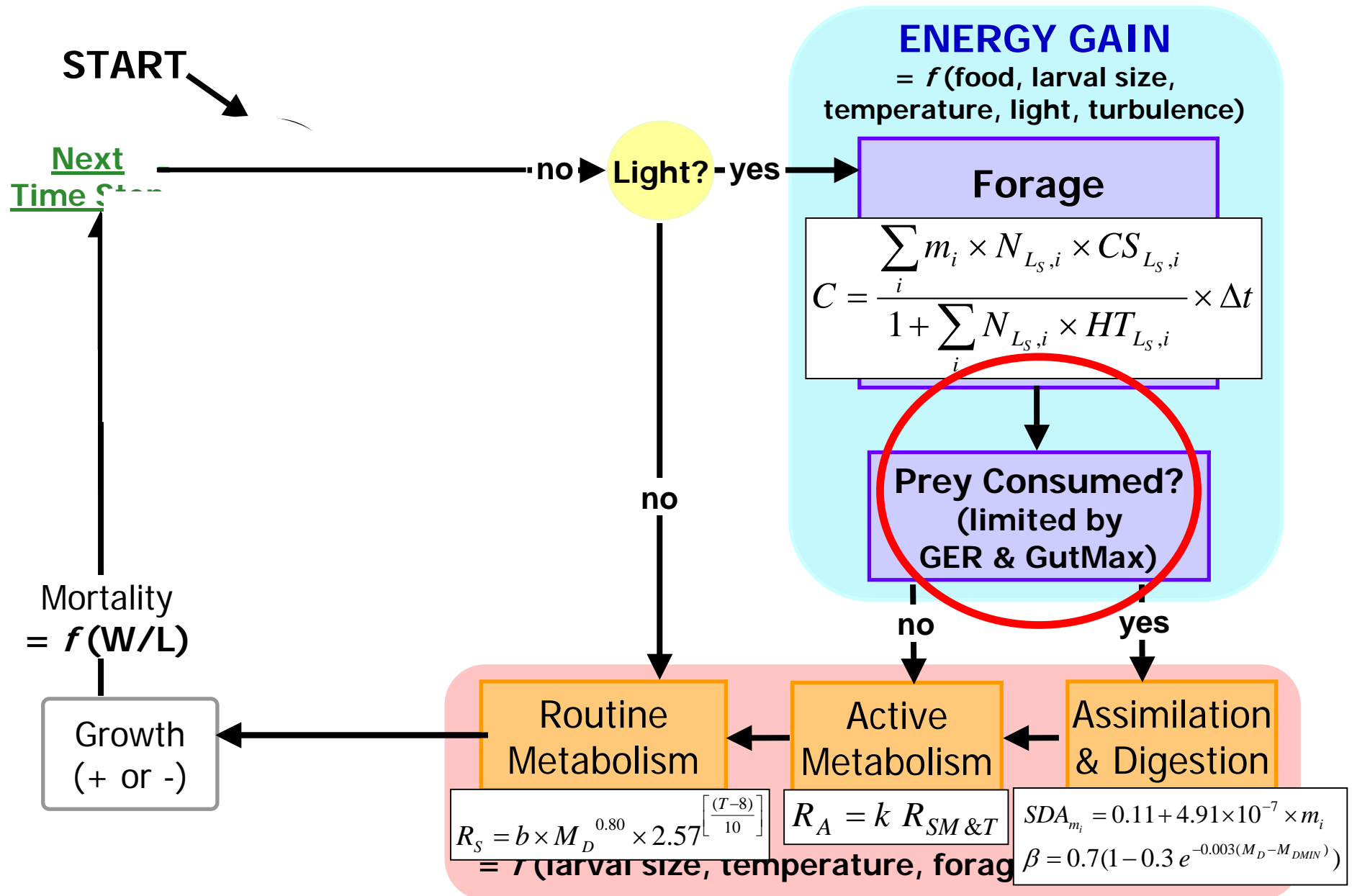


1-hr time step

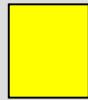
Rate of Energy Loss by Unfed Marine Fish Larvae (10 species, all rates adjusted to 8°C)



Mechanistic IBM: Foraging and Growth Subroutines



Rates of gut evacuation



Single meal



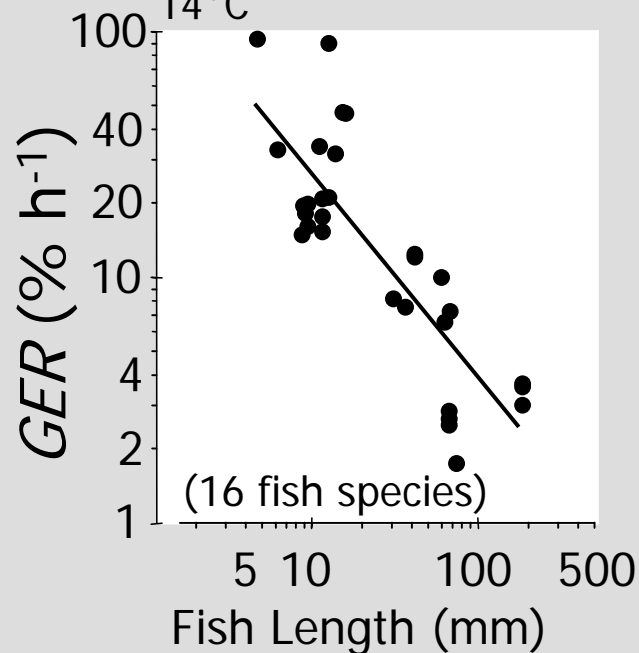
Constant feeding
 $CF = (2 \text{ to } 5) * SM$



Temperature
 $Q_{10} \sim 2.0$

$$GER = 1.79 * SL^{-0.83}$$

14°C

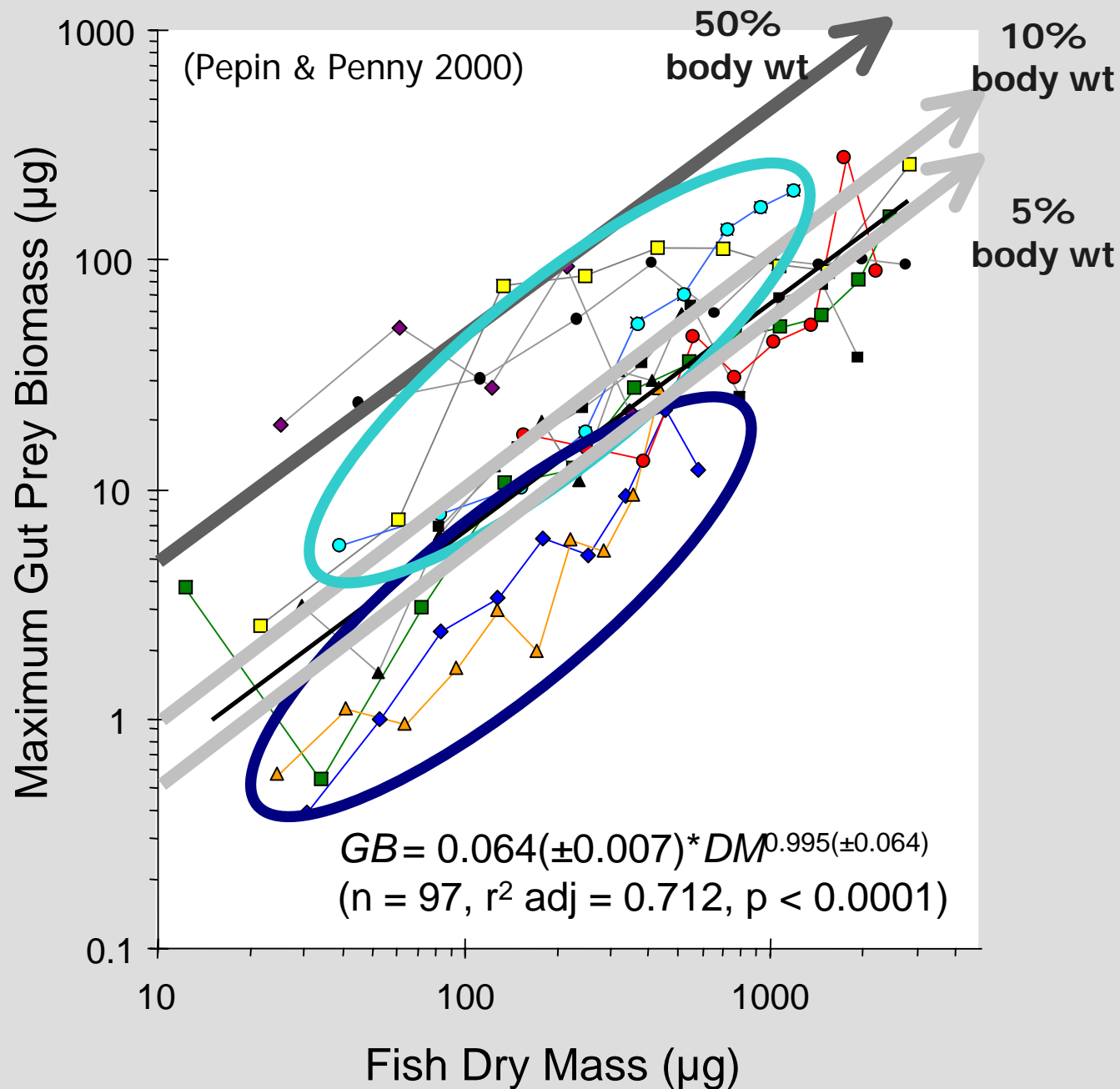


Species	Age (dph)	Length (min max)	T (°C)	Prey Conc. (# l ⁻¹)	Evacuation Rate single constant (% h ⁻¹)	Reference	
Marine Species (13)							
<i>Clupea harengus</i>	np	10 12	7	np	0.111	Blaxter 1962	
			11		0.200		
			15		0.222		
<i>Clupea harengus</i>	12	9 9	8	np	0.125	Blaxter 1965	
			15		0.250		
<i>Clupea harengus</i>	8-22	10.5 12	6-9	4-5*10 ⁸	0.667	Fossum 1983	
<i>Clupea harengus</i>	26-40	12.5 18.1	9.5	0.011-0.198	0.400	Pedersen 1984	
<i>Clupea harengus</i>	21-63	np np	9.2	3*10 ¹⁰ -10 ²	0.143	Werner & Blaxter 1979	
				3*10 ⁸	0.200-0.333		
				3*10 ⁴	0.250-0.170		
<i>Clupea harengus</i>		35 74	np	in situ	0.178	Arrhenius & Hansson 1994	
<i>Cynoscion regalis</i>	np	60 70	24	np	0.121-0.219	Lankford & Targett 1997	
<i>Gadus morhua</i>	7	(4-5)	5	np	0.500-0.667	Tilseth & Ellertsen 1984	
<i>Logadon rhomboids</i>				in situ	0.380	Peters and Kjelson 1975	
<i>Sardinops sagax</i>		10.1 13.9	20	in situ	0.500-0.250	Herrera & Balbontin 1983	
<i>Sebastes melanops</i>	np	35 93	7	ad libitum	0.019	Boehlert & Yoklavich 1983	
			12	ad libitum	0.029		
			18	ad libitum	0.039		
<i>Sprattus sprattus</i>	np	13 16	8-15	in situ	0.460	Peck et al. (unpubl. data)	
<i>Syngnathus fuscus</i>	np	150 200	15	np	0.038	Ryer & Boehlert 1983	
			23	np	0.078		
			27	np	0.107		
<i>Theragra chalcogramma</i>	< 7	(5.92)	6.2	1200 - 1500	0.207-0.246	Canino & Bailey 1995	
<i>Thunnus alalunga</i>	np	2.7 10	26	in situ	0.333-0.250	Young & Davis 1990	
<i>Thunnus maccoyii</i>	np	2.7 10	26	in situ	0.333-0.250	Young & Davis 1990	
<i>Trachurus declivis</i>	np	2.4 14.3	15-18	in situ	0.167 - 0.25	Young & Davis 1992	
<i>Ulvaria subbiturvata</i>	np	4 13	14	in situ	0.165-0.290	Bochdansky et al. In Press	
Freshwater Species (9)							
<i>Cyprinus carpio</i>	np	8 12	18-29		0.050	0.125-1.000	Chiba 1961
<i>Coregonus albula</i>	14	8.7 18		np	0.280		Karjalainen et al. 1991
<i>Dorosoma cepedianum</i>	np	25 89	21	np	0.130-0.250	0.550-1.250	Shepherd & Mills 1996
<i>Micropterus salmoides</i>	np	20 60*	18	np	0.192	0.357	Laurence 1971
			23		0.263	0.500	
<i>Perca Flavescens</i>	np	(17-19.5)	21	np	np	1.667	Noble 1973
		30-40	22	np	0.154	0.667	
		(60)	15	np	0.083	0.167	
<i>Perca Flavescens</i>	np	20 69	14-21	np	np	0.417-3.333	Mills et al. 1984
<i>Perca fluviatilis</i>	np	(13.1)	np	field	0.400		Worischka & Mehner 1998
<i>Salmo salar</i>	np	43 99	9-13	np	0.017	0.068	Talbot et al. 1984
<i>Stizostedion lucioperca</i>	np	10.6	np	field	0.430		Worischka & Mehner 1998
<i>Stizostedion vitreum</i>	np	10.4 16.2	15	np	0.109		Johnston & Mathias 1996
			20	np	0.245		
			25	np	0.106		
<i>Stizostedion vitreum</i>	21	(29.4)	22	np	0.167	0.500	Corazza & Nickum 1983

*estimate based upon range in dry weights (196 to 721 mg)

Amount of Prey in Gut Contents

- ◆ *Clupea harengus*
- *Hippoglossoides platessoides*
- ▲ *Mallotus villosus*
- *Pleuronectes ferrugineus*
- *Stichaeus punctatus*
- ◆ *Pleuronectes americanus*
- *Ulvaria subbifurcata*
- *Liparus sp.*
- *Gadus morhua*
- ▲ *Glyptocephalus cynoglossus*
- all species



1-D to 3-D Modelling of potential survival & growth (gadoid vs clupeid larvae)

Temperature-dependent energy loss
(daily, active metabolism, etc.)

vs.

Temperature-dependent energy gain
(prey size, gut evacuation & gut biomass, etc.)



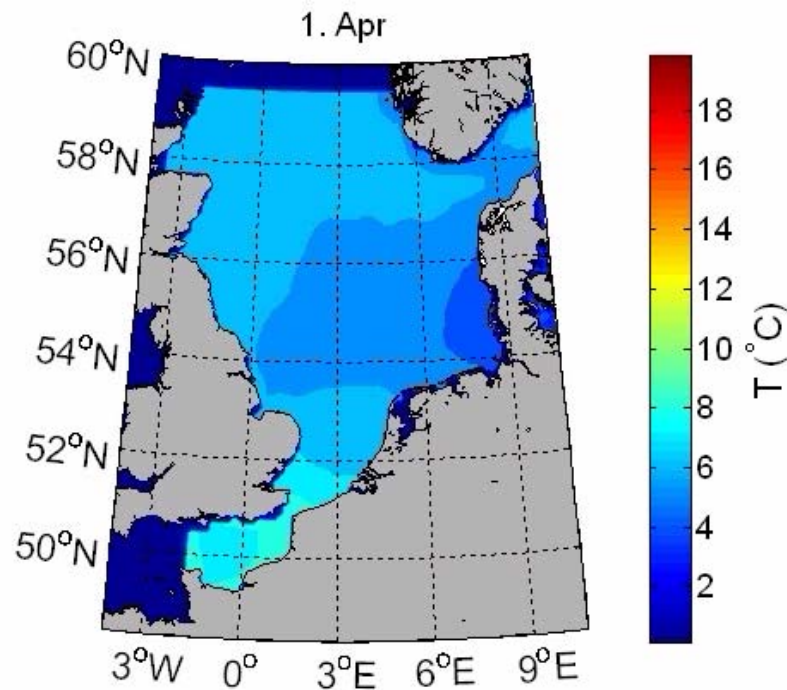
Atlantic Cod



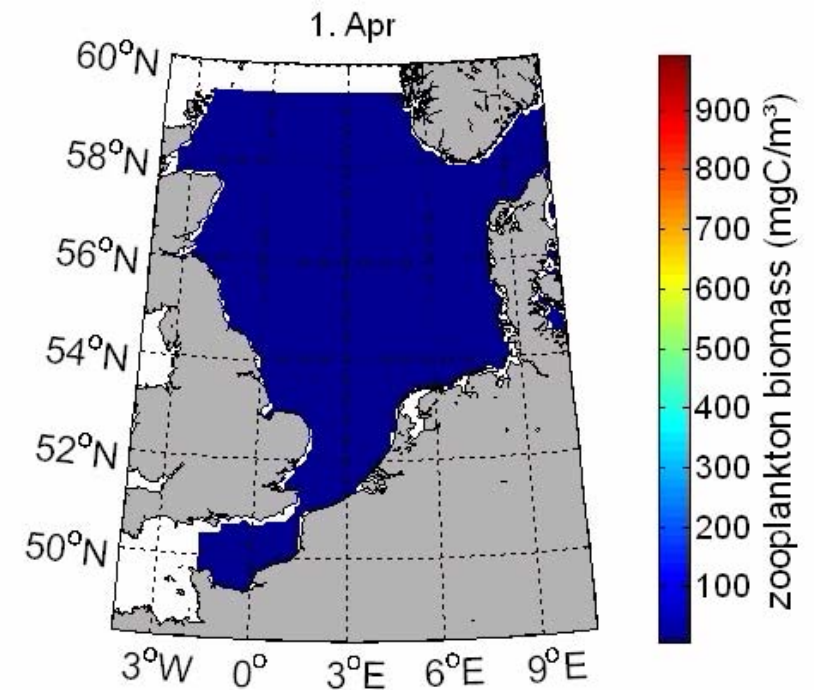
Sprat

3-D coupled model system (NPZD component)

Temperature



Seasonal Zooplankton Dynamics



Year = **1993**
all values
depth-averaged

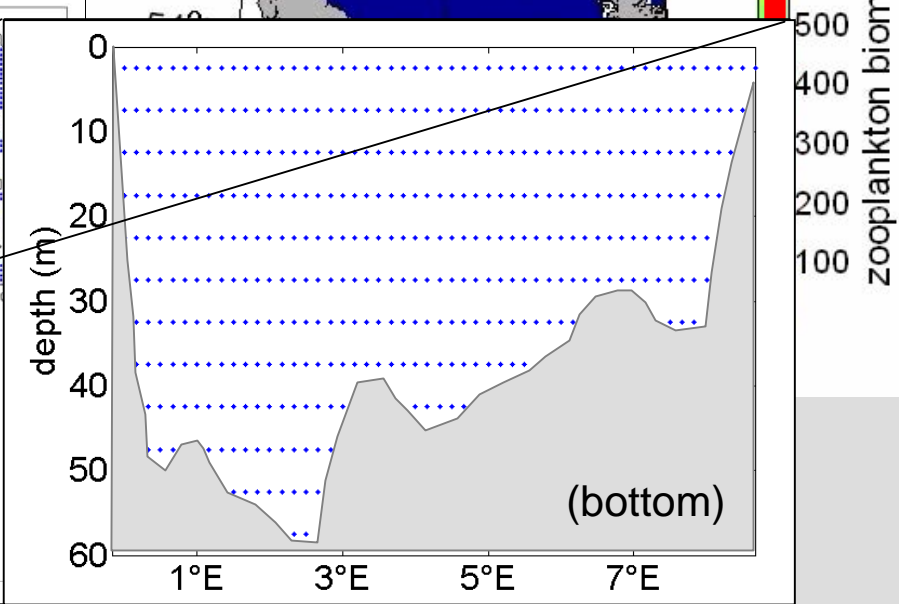
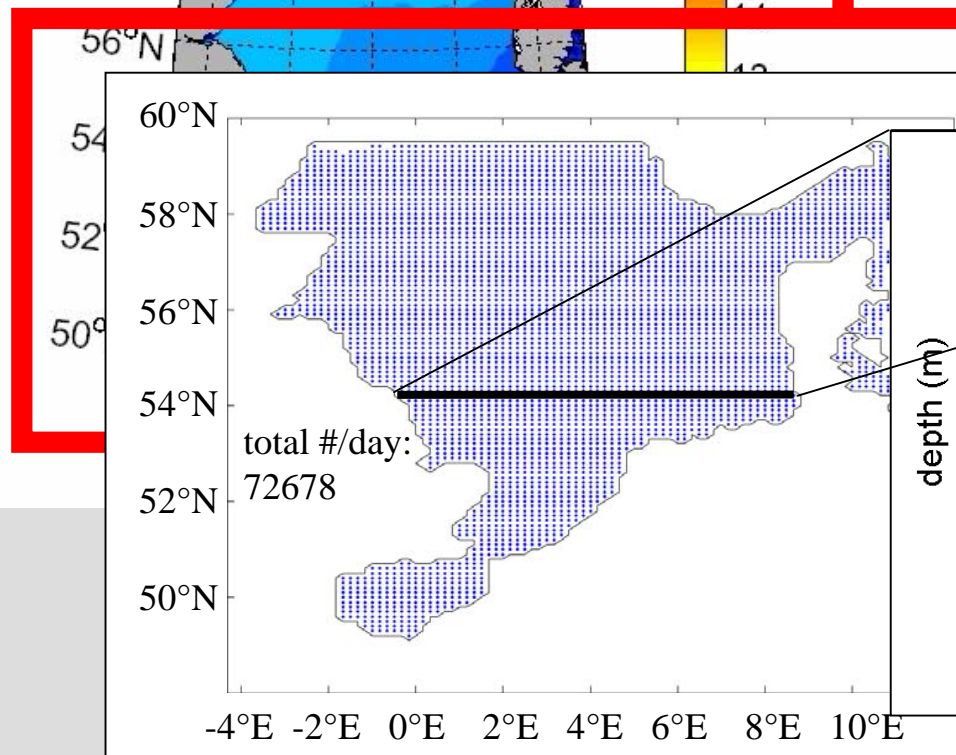
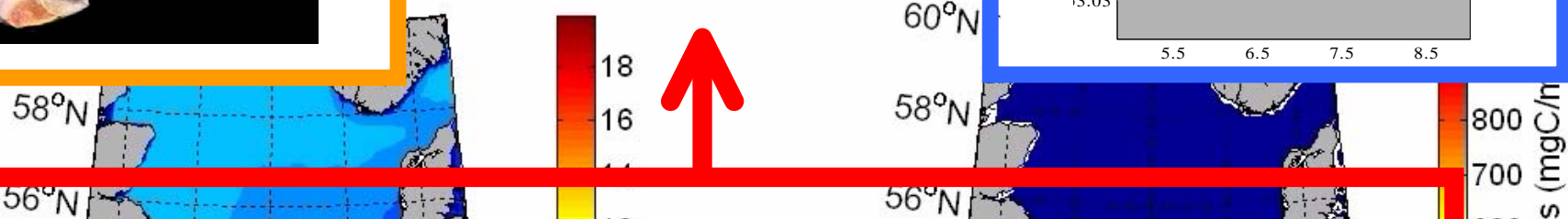
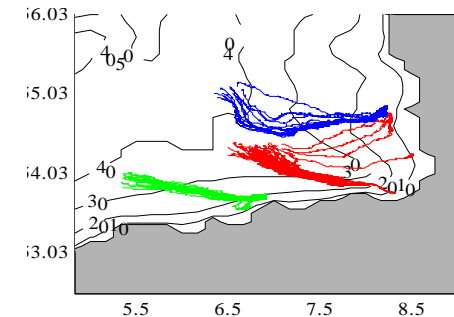
Ecosystem Model
2 Phytoplankton groups
2 Zooplankton groups
3 Nutrient cycles

mechanistic IBM
for larval fish



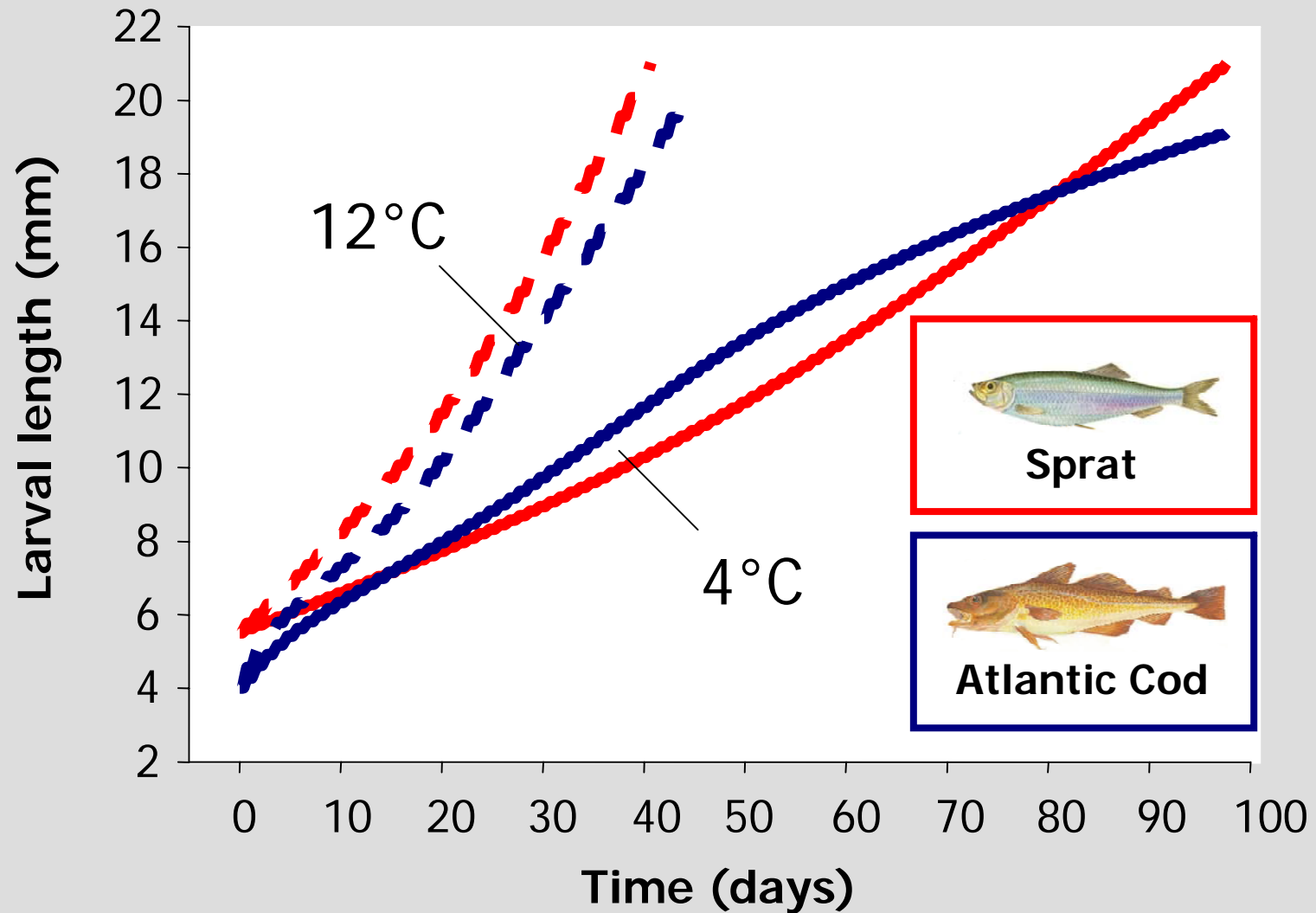
3-D Coupled model system

3D circulation &
particle tracking



But, first some simple 1-D results....

Larval growth when prey is unlimited



1-D model) temperature-specific prey requirements

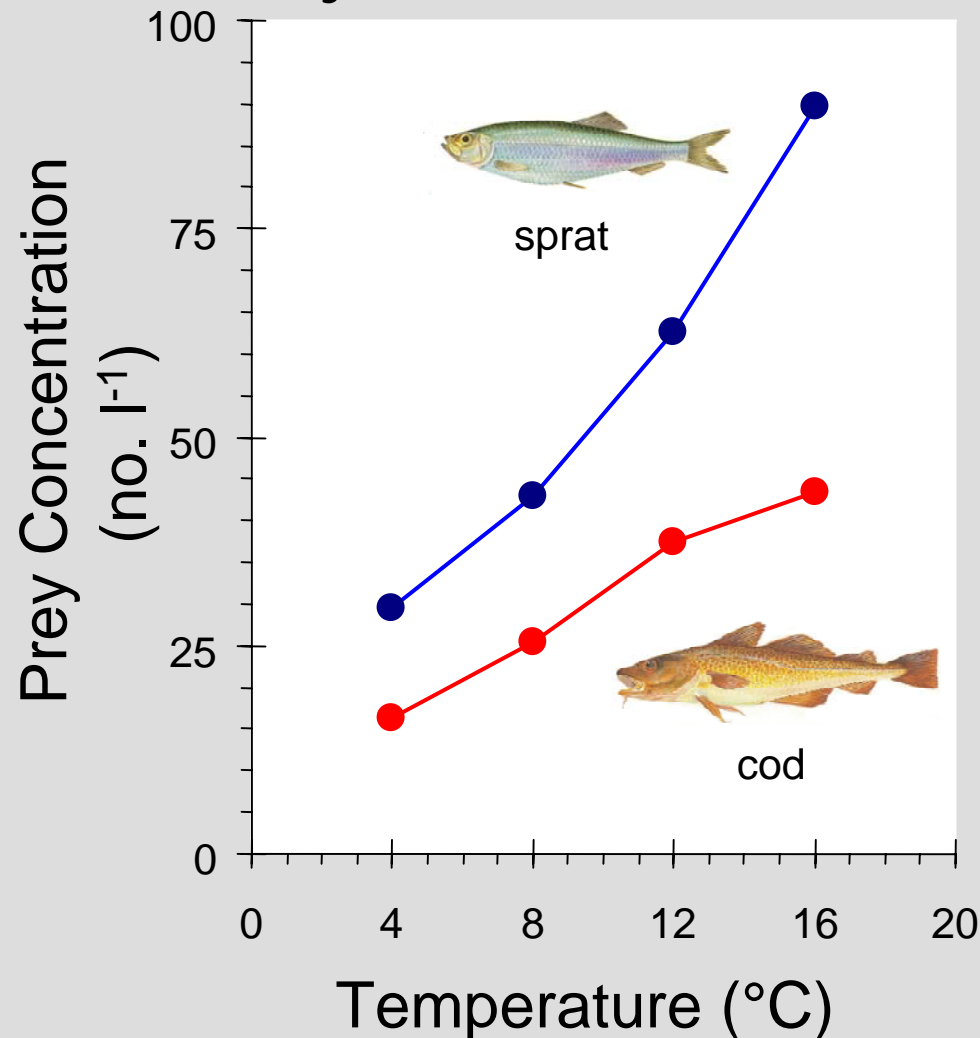


First-feeding
150 - 700 μm prey



First-feeding
150 - 300 μm prey

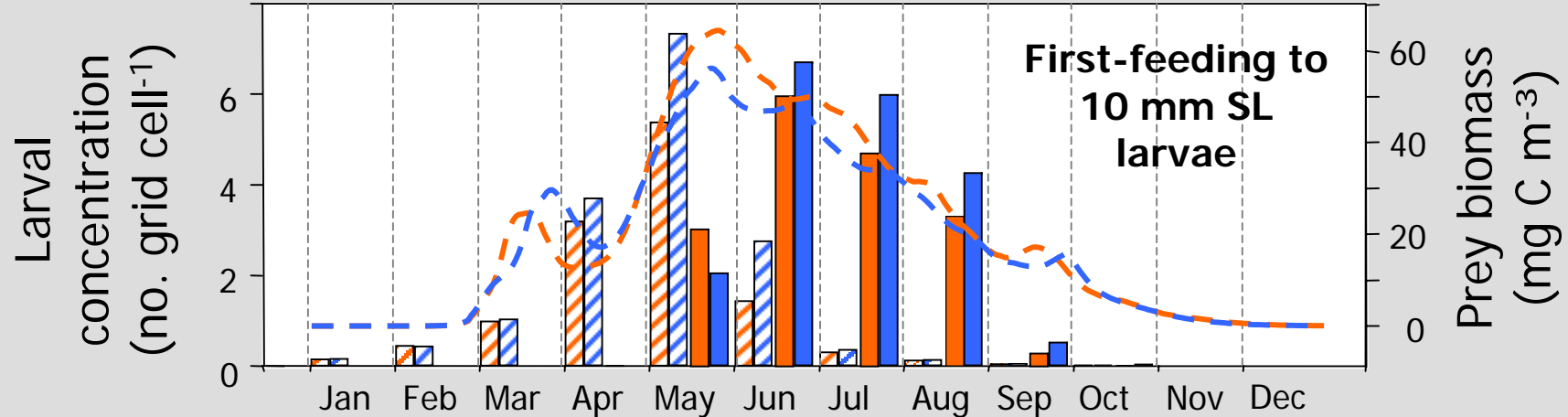
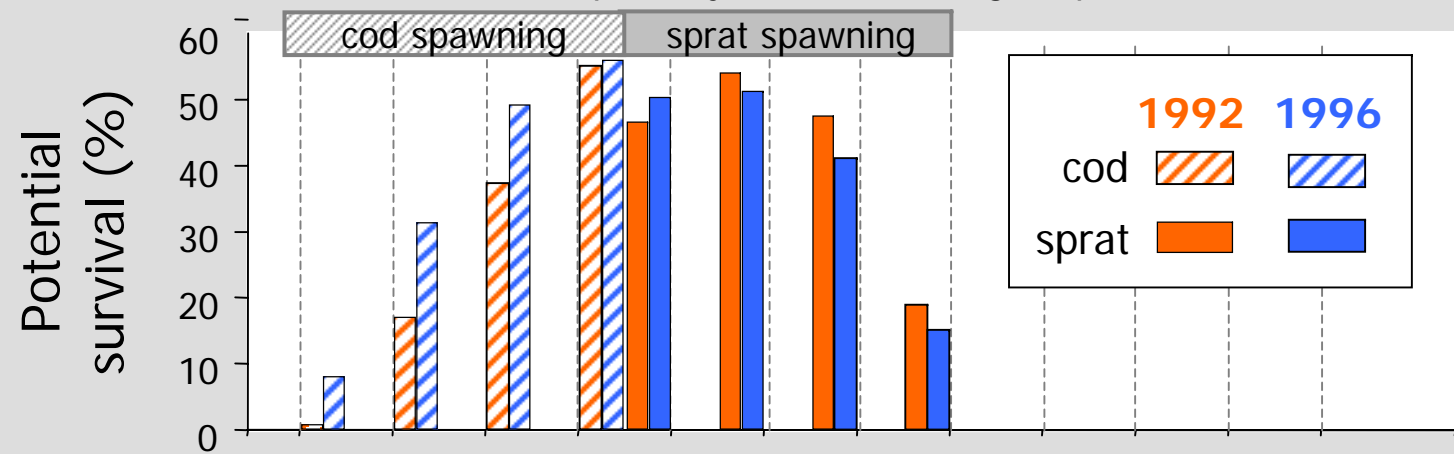
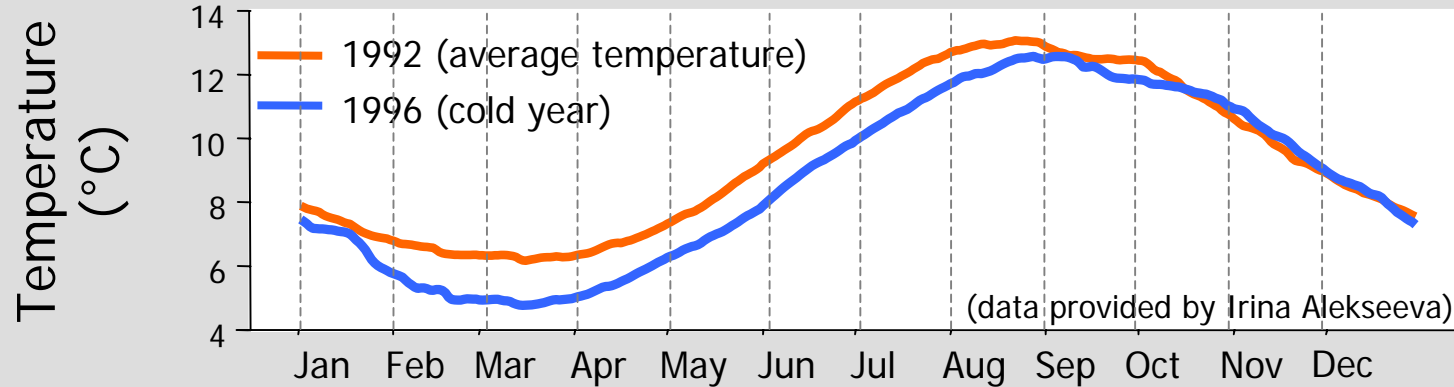
Prey Threshold for Survival



**Fish don't eat
carbon...**

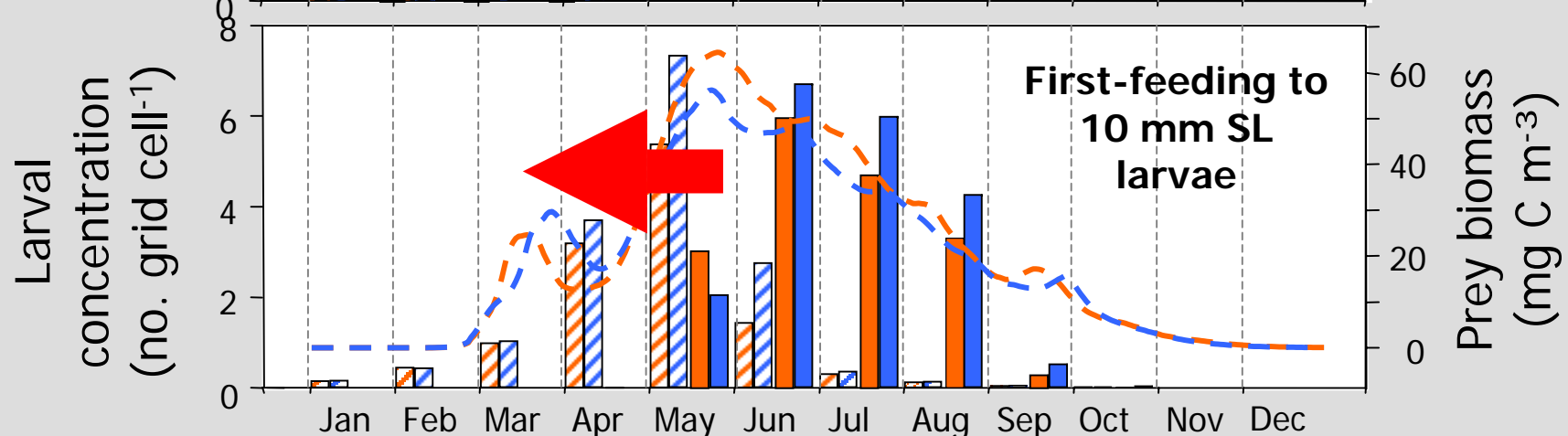
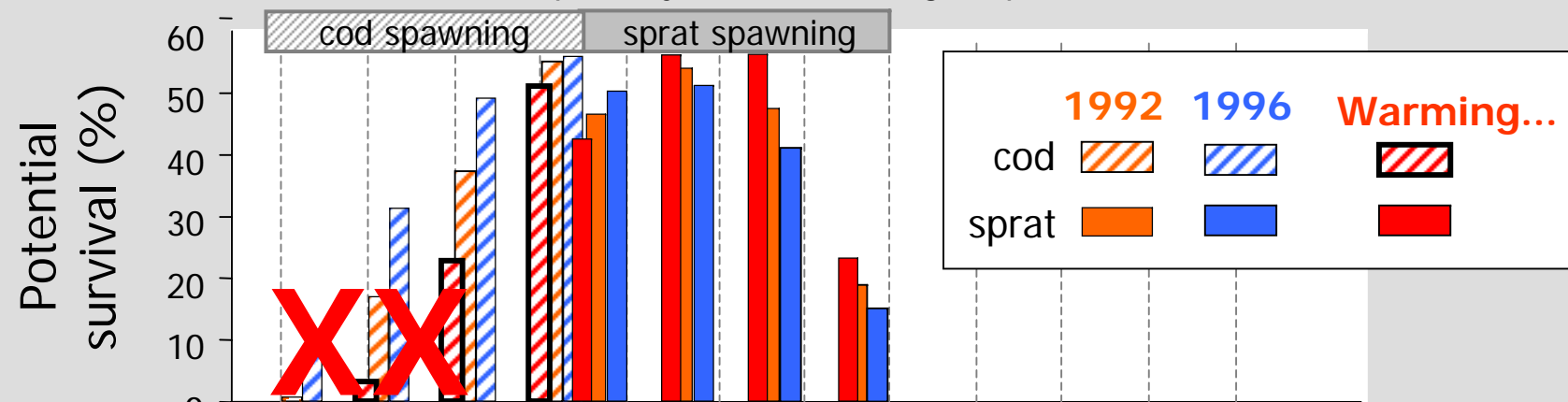
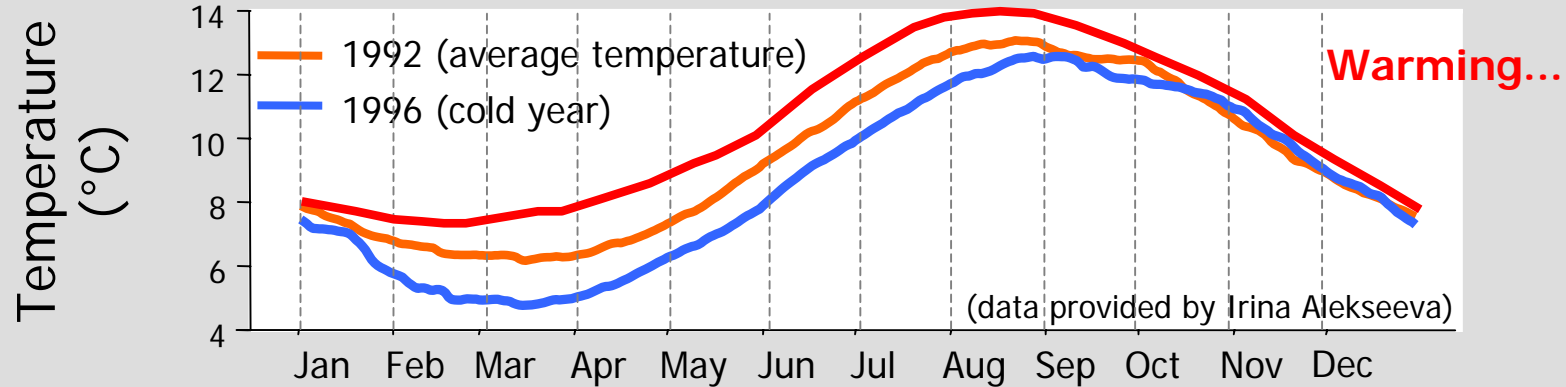
**(converted to
copepods in size
classes
available to first
feeding larvae)**

3-D Coupled Model Results: North Sea Cod & Sprat



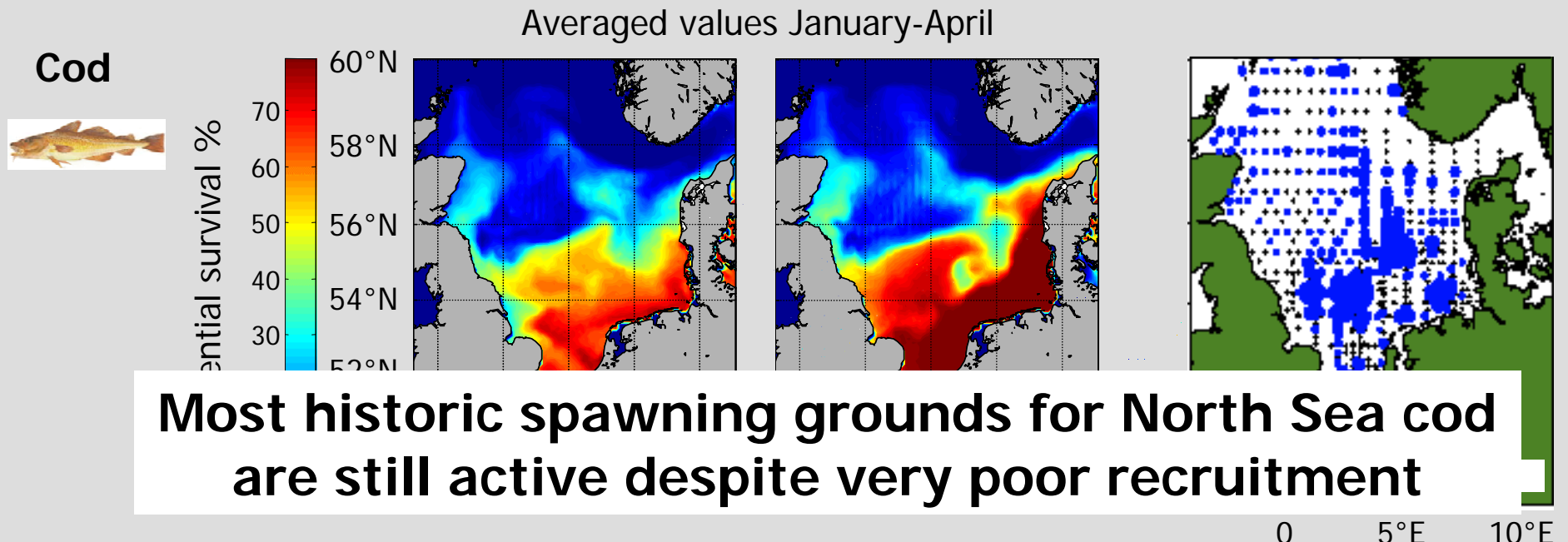
Time (accounting for drift of eggs & larvae) (Daewel et al. In prep)

3-D Model Results: North Sea Cod and sprat

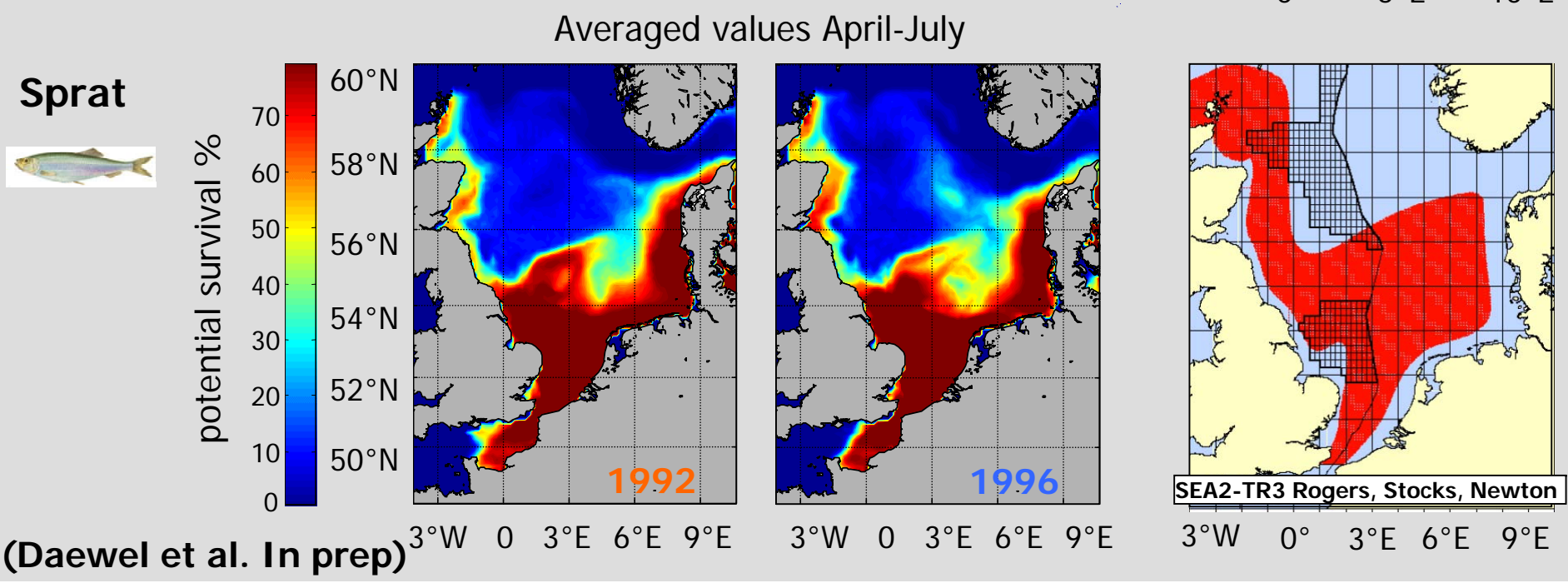


Time (accounting for drift phase) (Daewel et al. In prep)

3d coupled-model: areas supporting potential survival compared to observations



Most historic spawning grounds for North Sea cod are still active despite very poor recruitment



(Daewel et al. In prep)

Summary & Conclusions I

Considerable variation exists in physiological attributes of marine fish early life stages that are directly impacted by climate – examples:

Eggs

Change in Development Rate vs Temperature

Yolksac Larvae

Change in size-at-hatch vs temperature

Change in the time to end of yolk reserves vs temperature

Feeding Larvae (inter-specific differences)

rate of energy loss when unfed

maximum biomass of prey in gut & prey size vs body size

Examine interlinkages among physiological traits & rates that change with body size &/or temperature

$$(C = G + R + E + F)$$

Summary & Conclusions II

Biophysical IBMs can test how species-specific differences in physiological traits may influence severity of climate impacts on early life stage vital rates (e.g., cod vs sprat in North Sea)

Growth response & temperature

Similar in larval cod & sprat when provided unlimited prey

Prey thresholds for survival & growth

Differ between cod & sprat (cod has larger prey field + higher max. gut content, and species have similar energy losses)

Match-mismatch and warming

Fidelity to spawning site & time may lead to severe mismatch between cod larvae & zooplankton production. Sprat appears much less vulnerable to warming.

Work is ongoing with coupled NPZD-IBM ...

(e.g., inter-annual (20+ yr) comparison for both species)

Acknowledgements

Data Sets

Franziska Bils
Christian Lindemann
Christof Schneider
Audrey Geffen
Christopher Chambers
Greg Lough:

Co-authors

Ute Daewel (hard at work)
Corinna Schrum



Thanks for your Attention!

Questions?, Comments... .. Lunch?

